

$R_1, R_2 = H, \text{ alkyl, aryl, } -OH, -COOH, -X, -NH_2, \text{ arylacyl, hydroxyacyl, carboxyacyl, aminoacyl, ketoacyl, halogenated acyl, ester} \dots$

Continued from
Figure 1A

Continued from
Figure 1A

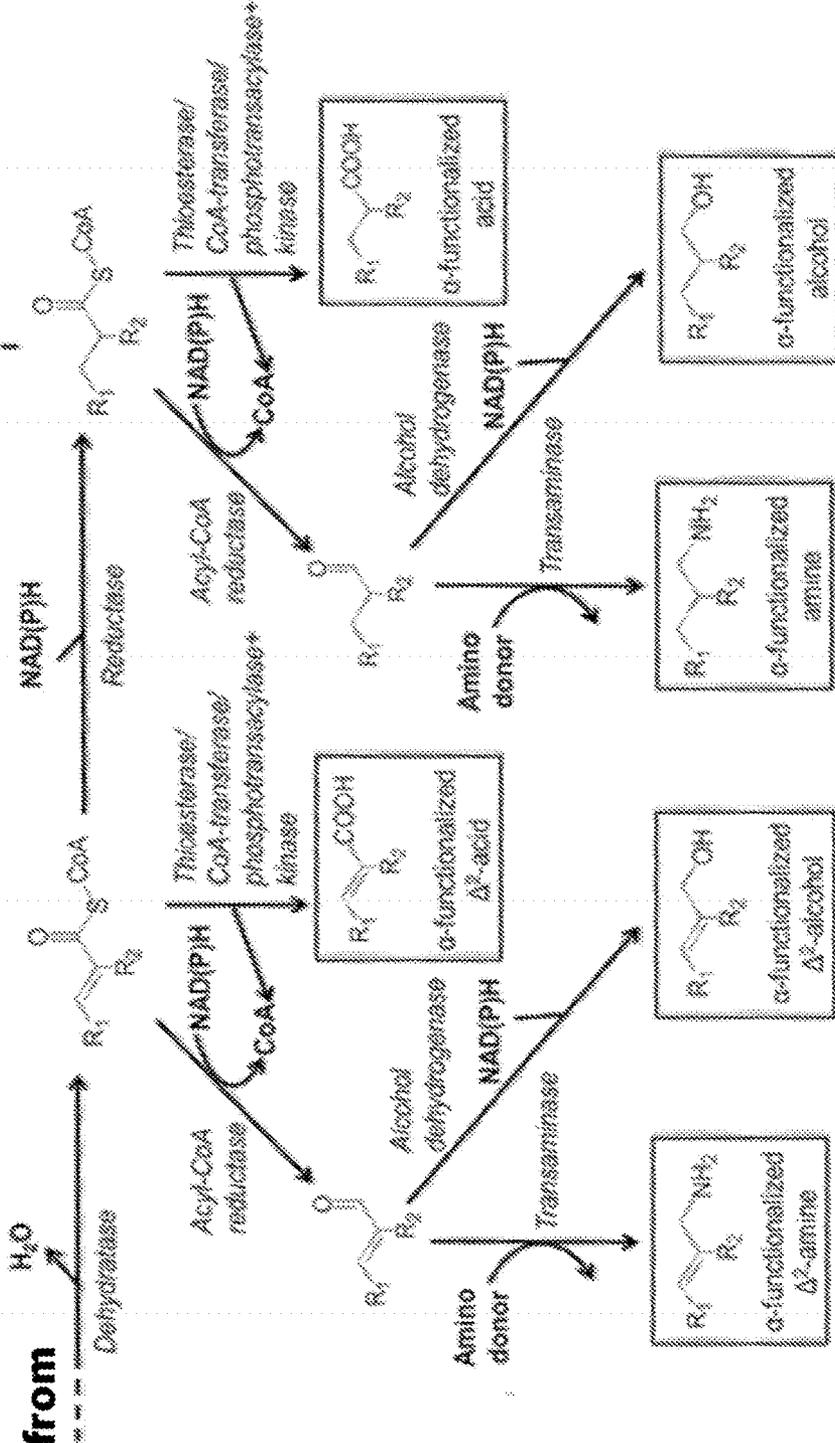


Figure 1B

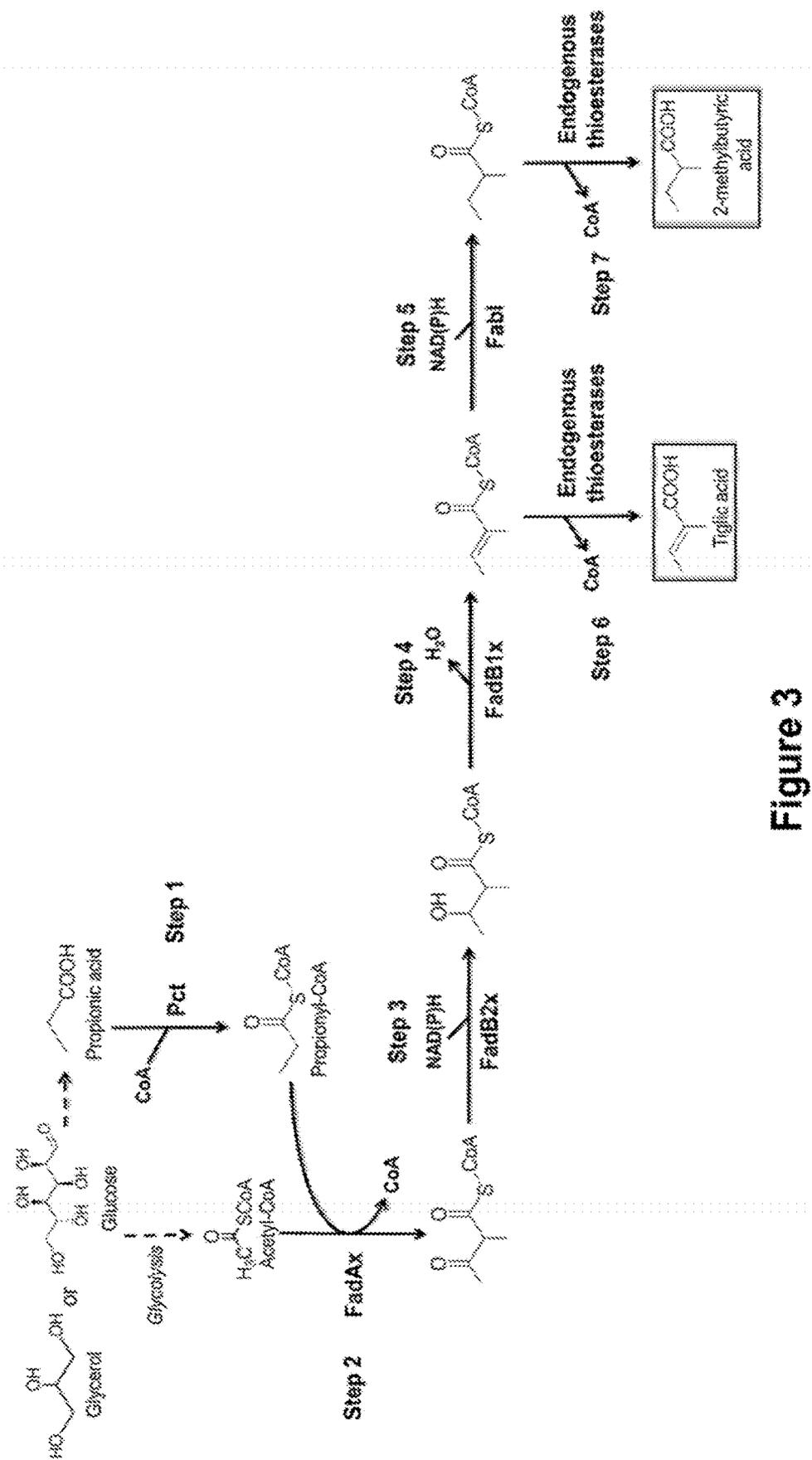


Figure 3

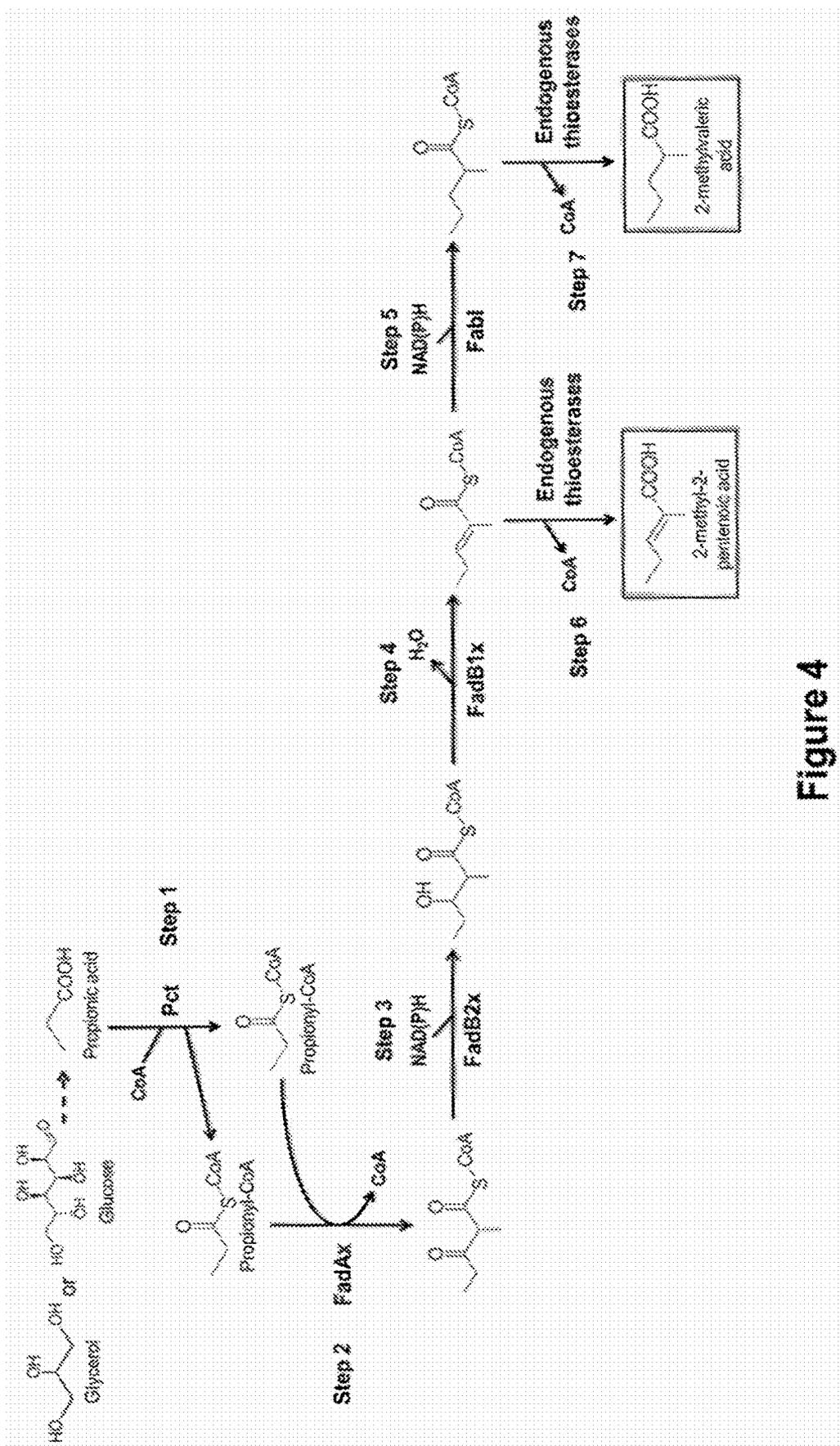


Figure 4

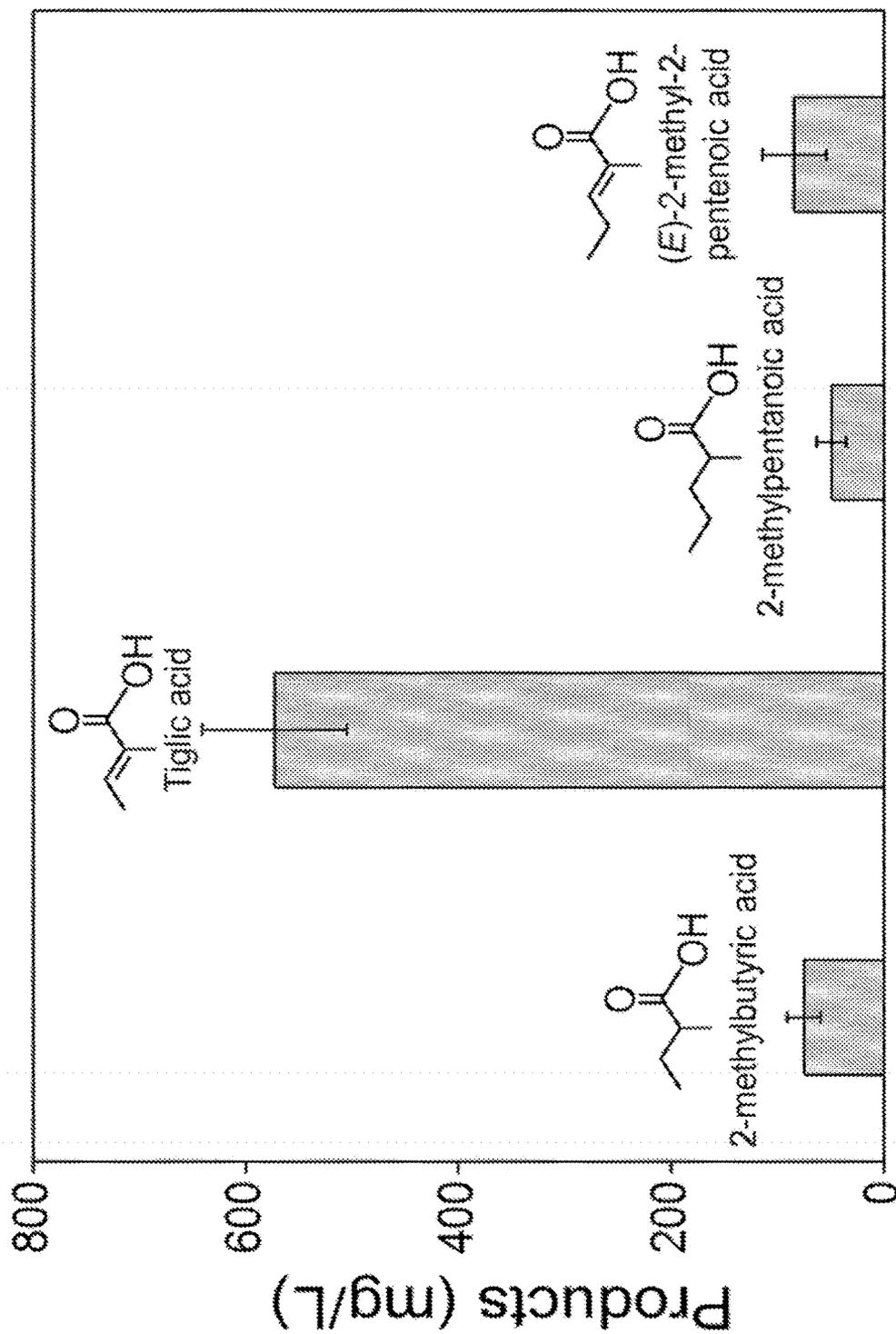


Figure 5

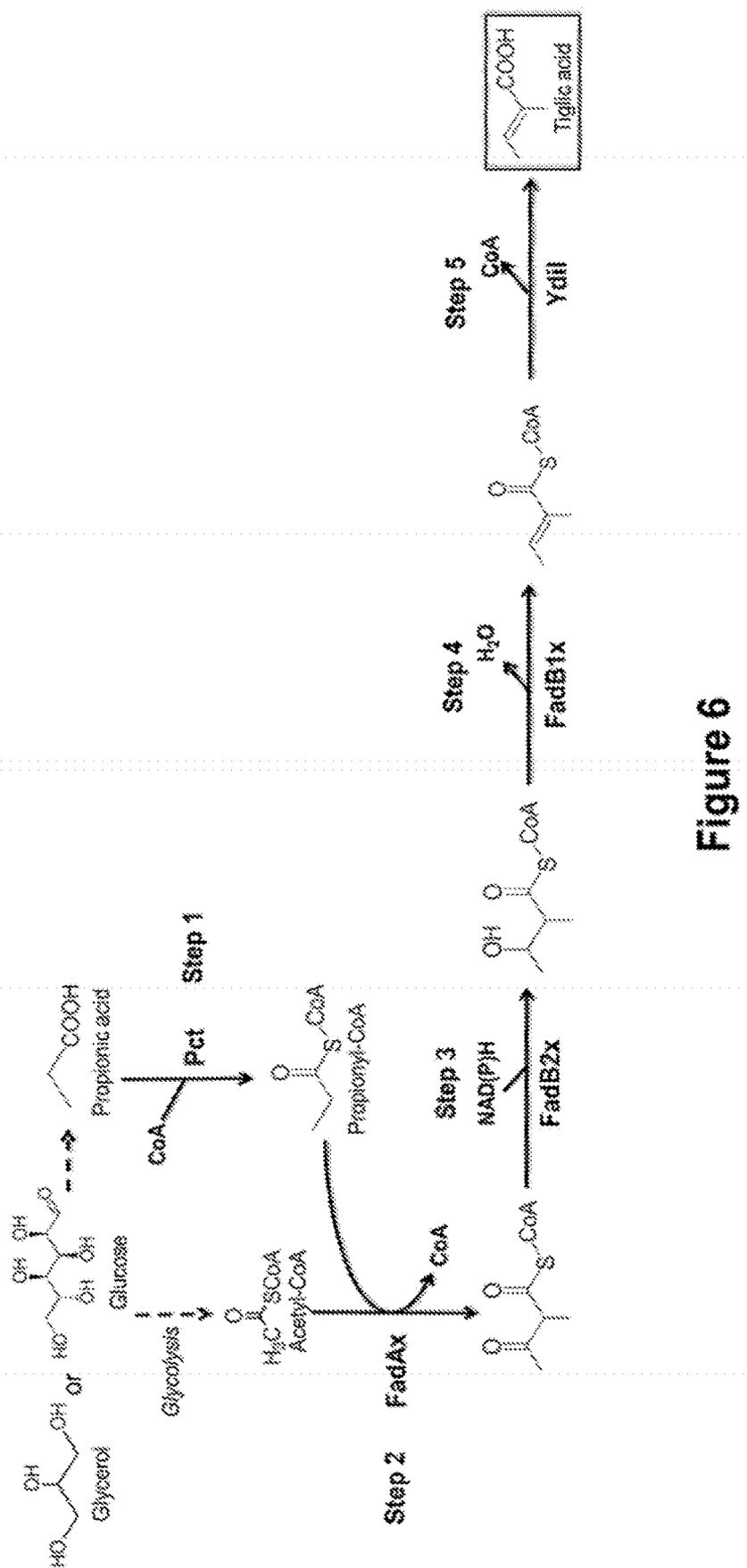


Figure 6

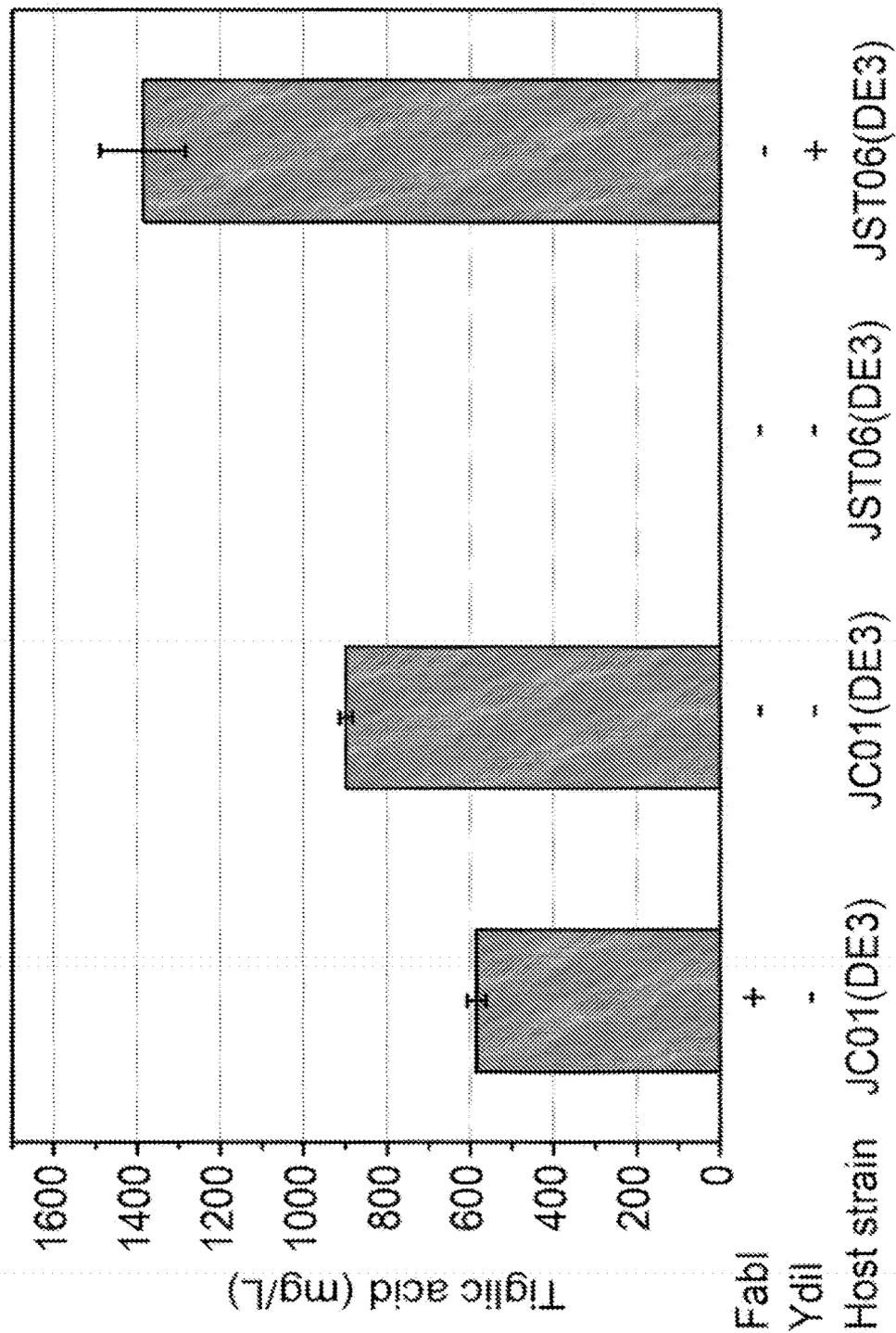


Figure 7

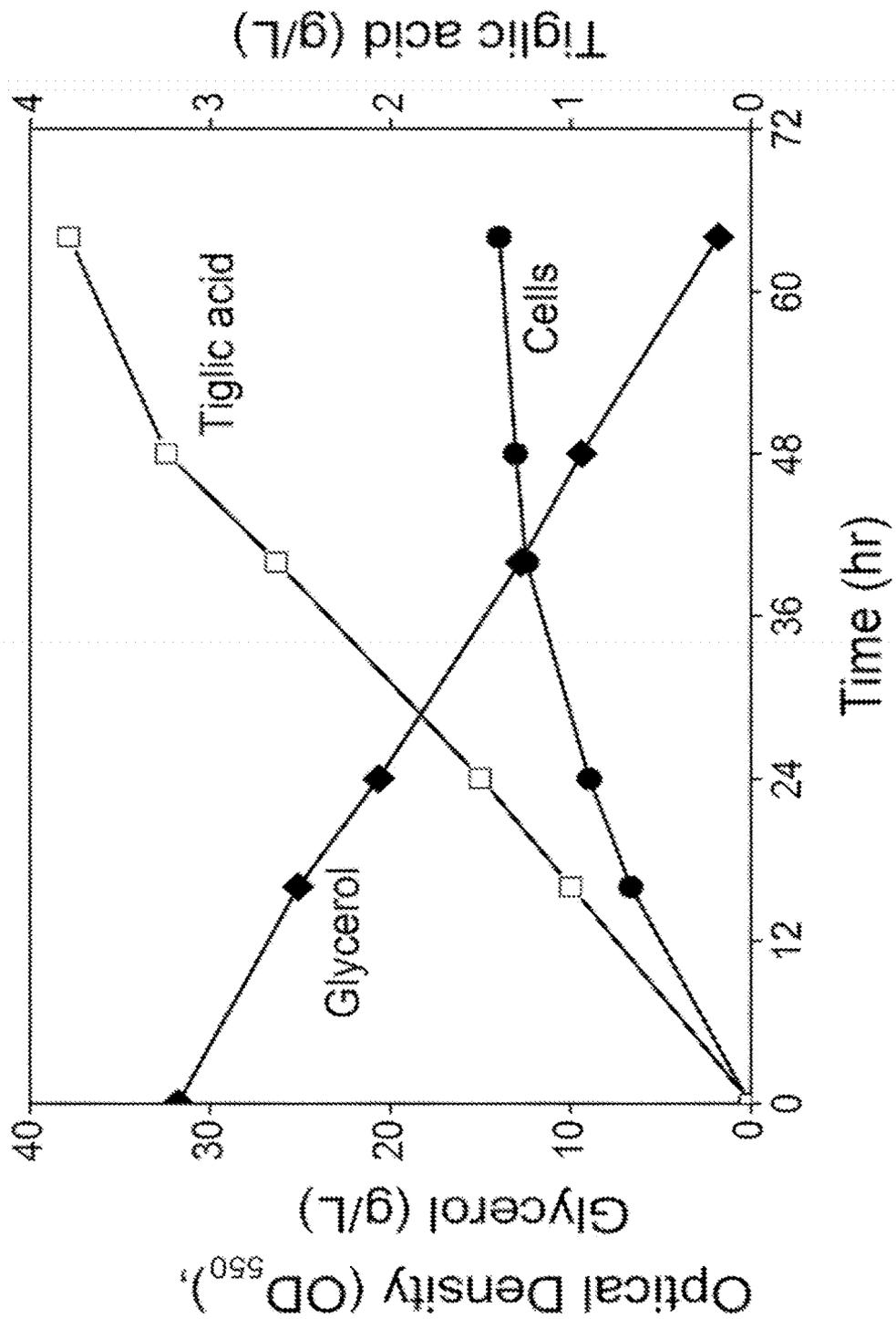


Figure 8

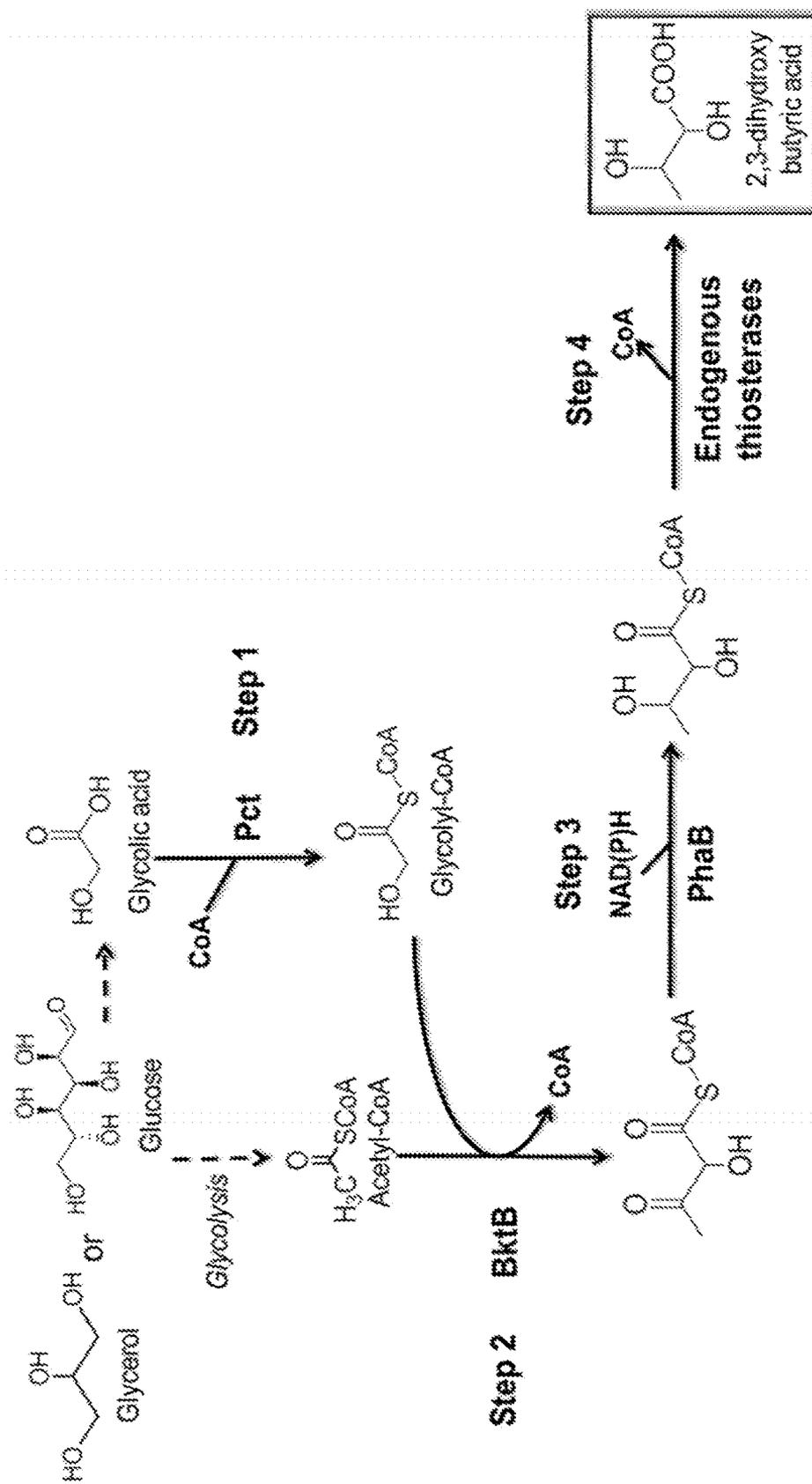


Figure 10

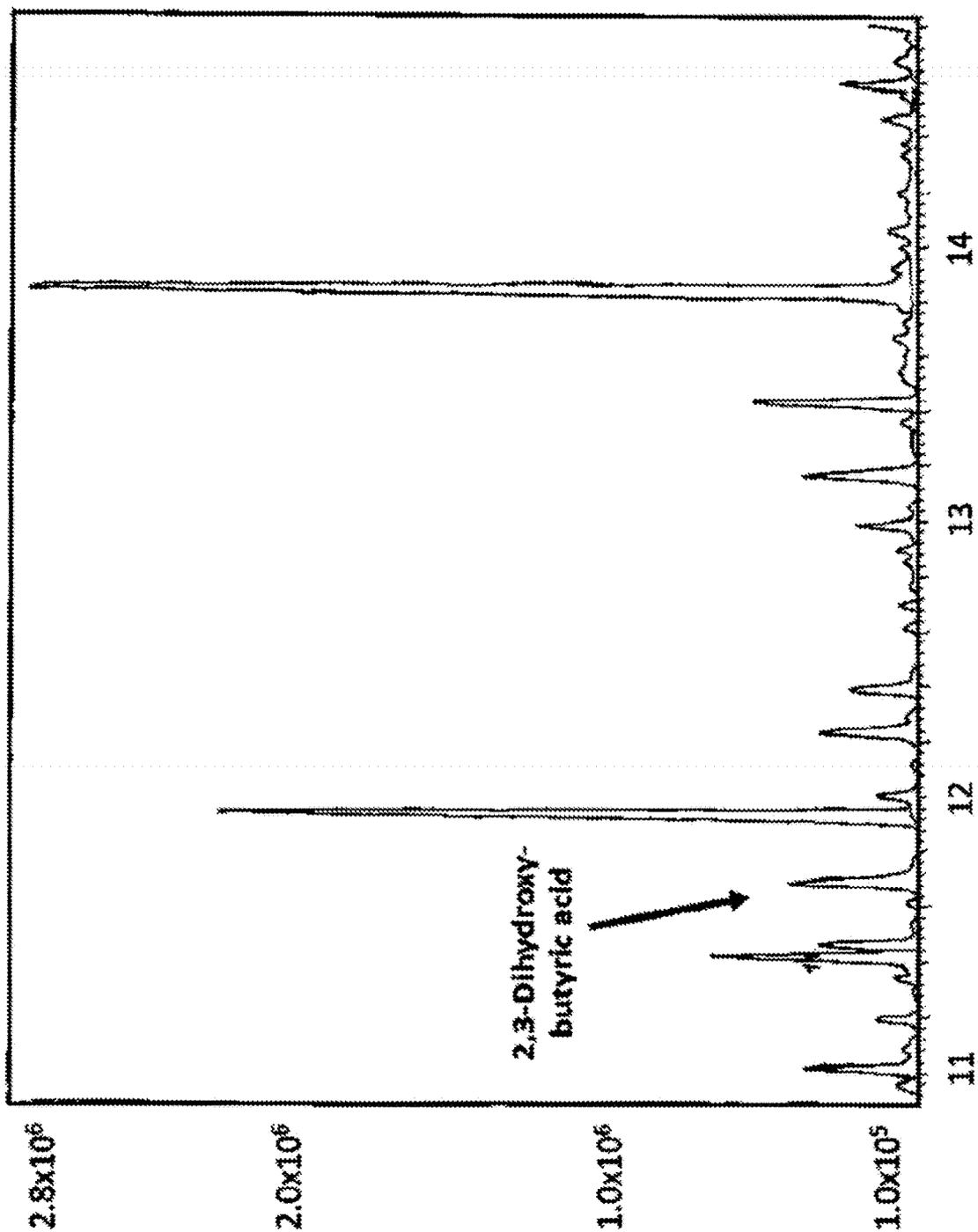


Figure 11

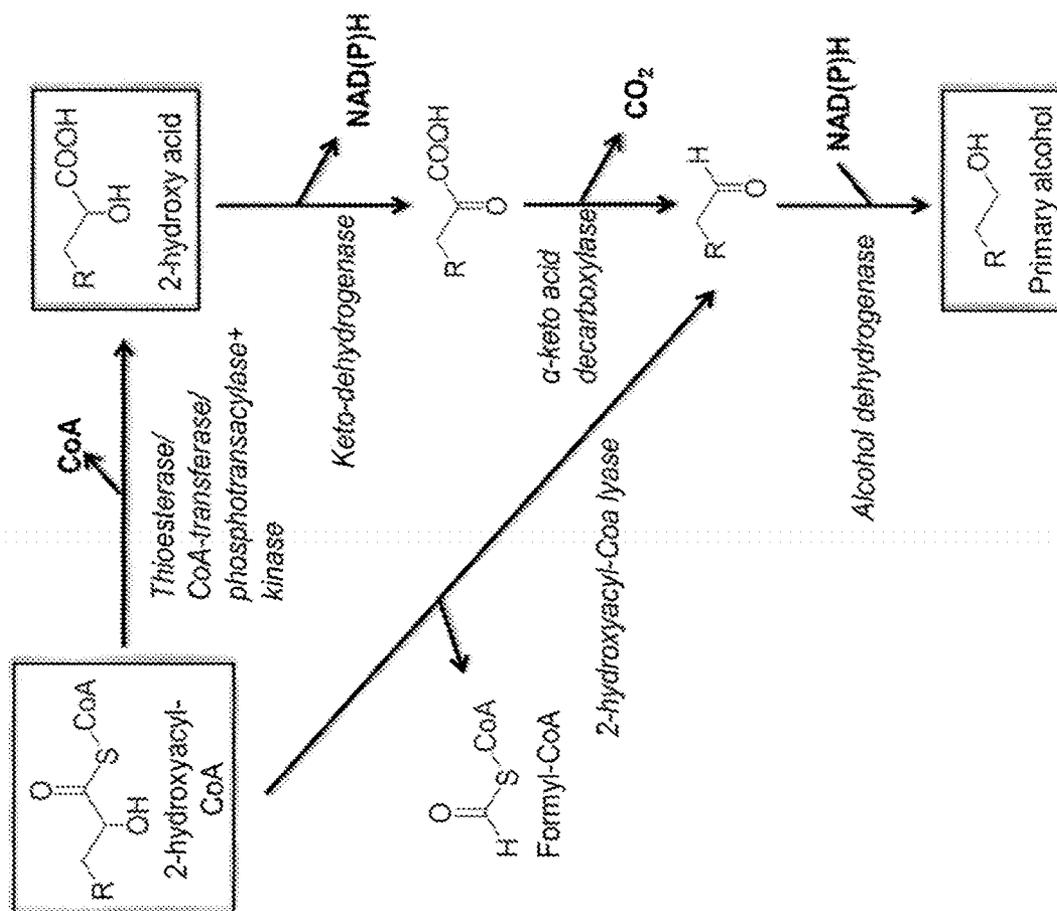


Figure 12

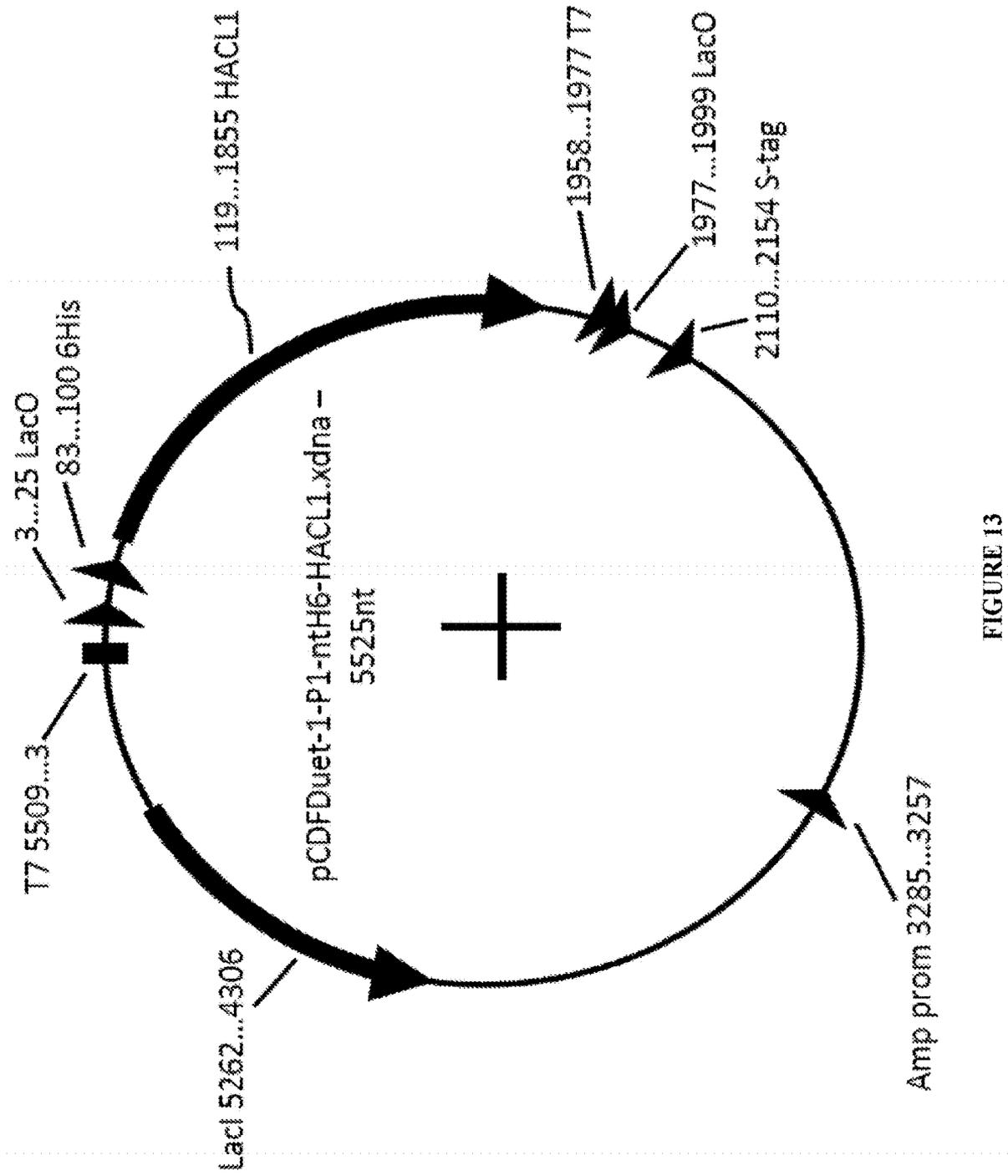
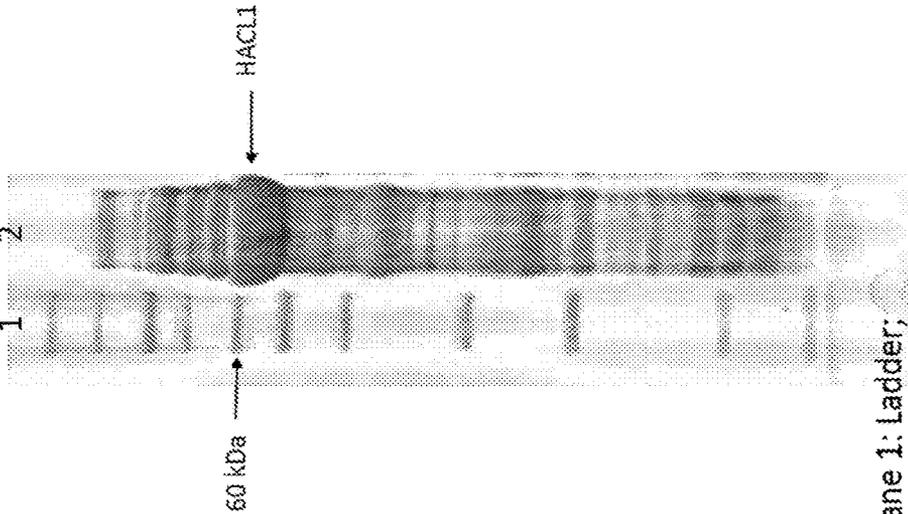


FIGURE 13

Expression of *Homo sapiens* HACL1 in the whole cell fraction of BL21(DE3)



Lane 1: Ladder;
Lane 2: Whole cell sample of BL21(DE3) pCDFDuet-1-P1-nth6-HACL1

Figure 14

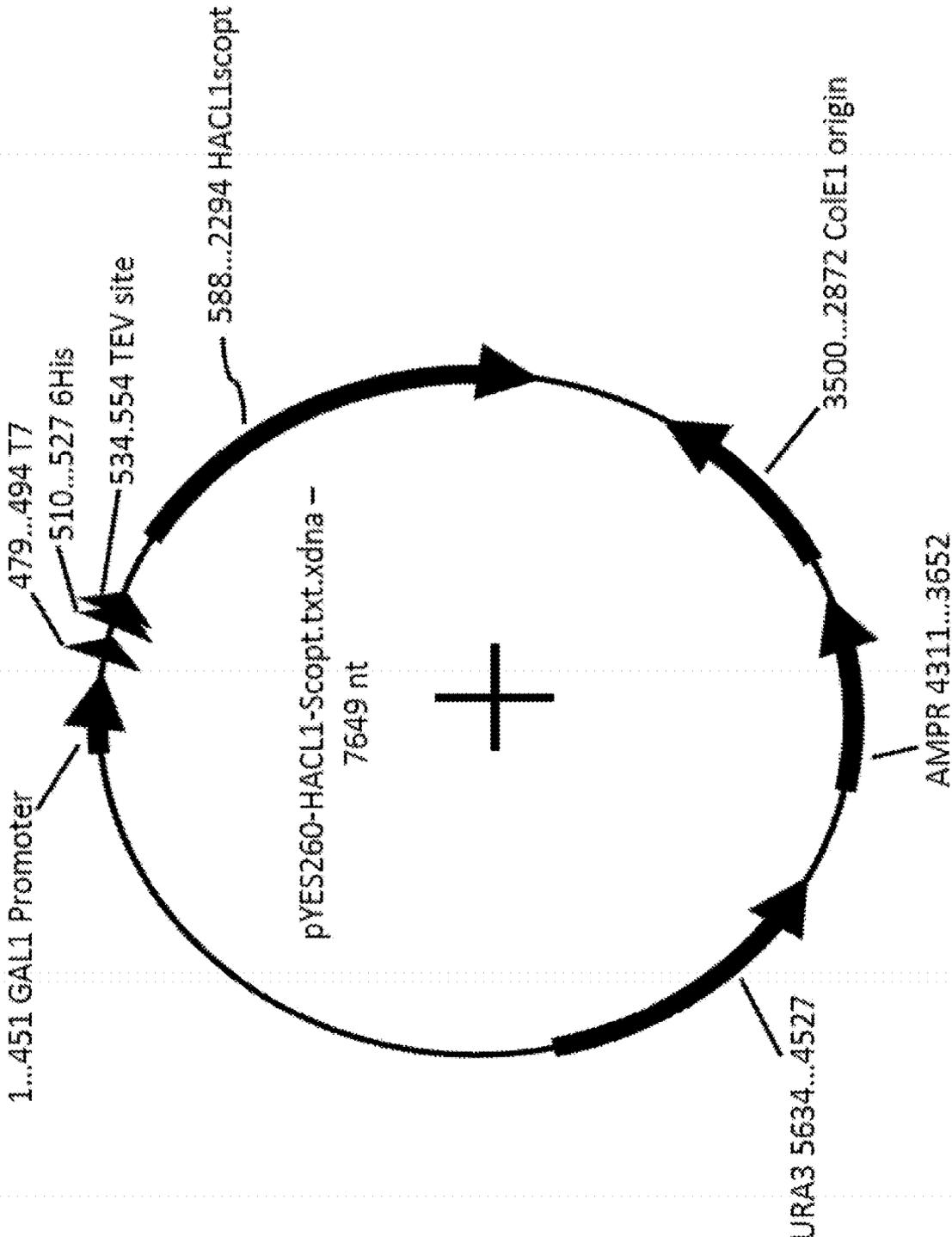
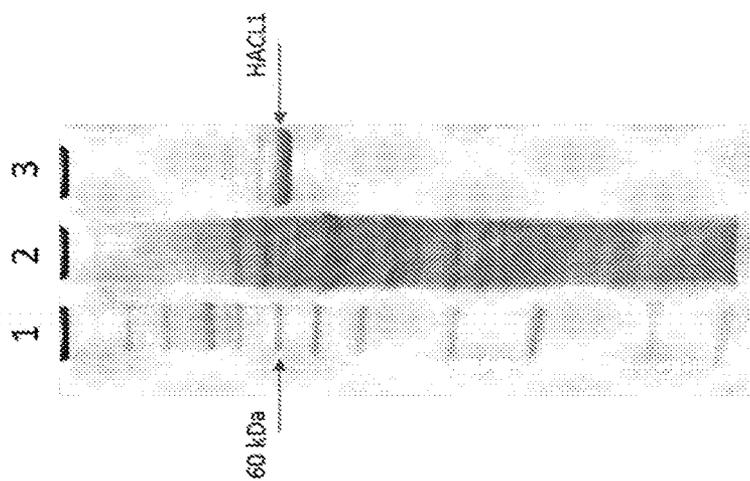


FIGURE 15

Expression and purification of *Homo sapiens*
HACL1 in *S. cerevisiae* INVSc1



Lane 1: Ladder;
Lane 2: Cell extract fraction of *S. cerevisiae*
pYES260-HACL1-ScOpt;
Lane 3: HACL1 purified from *S. cerevisiae*
pYES260-HACL1-ScOpt

Figure 16

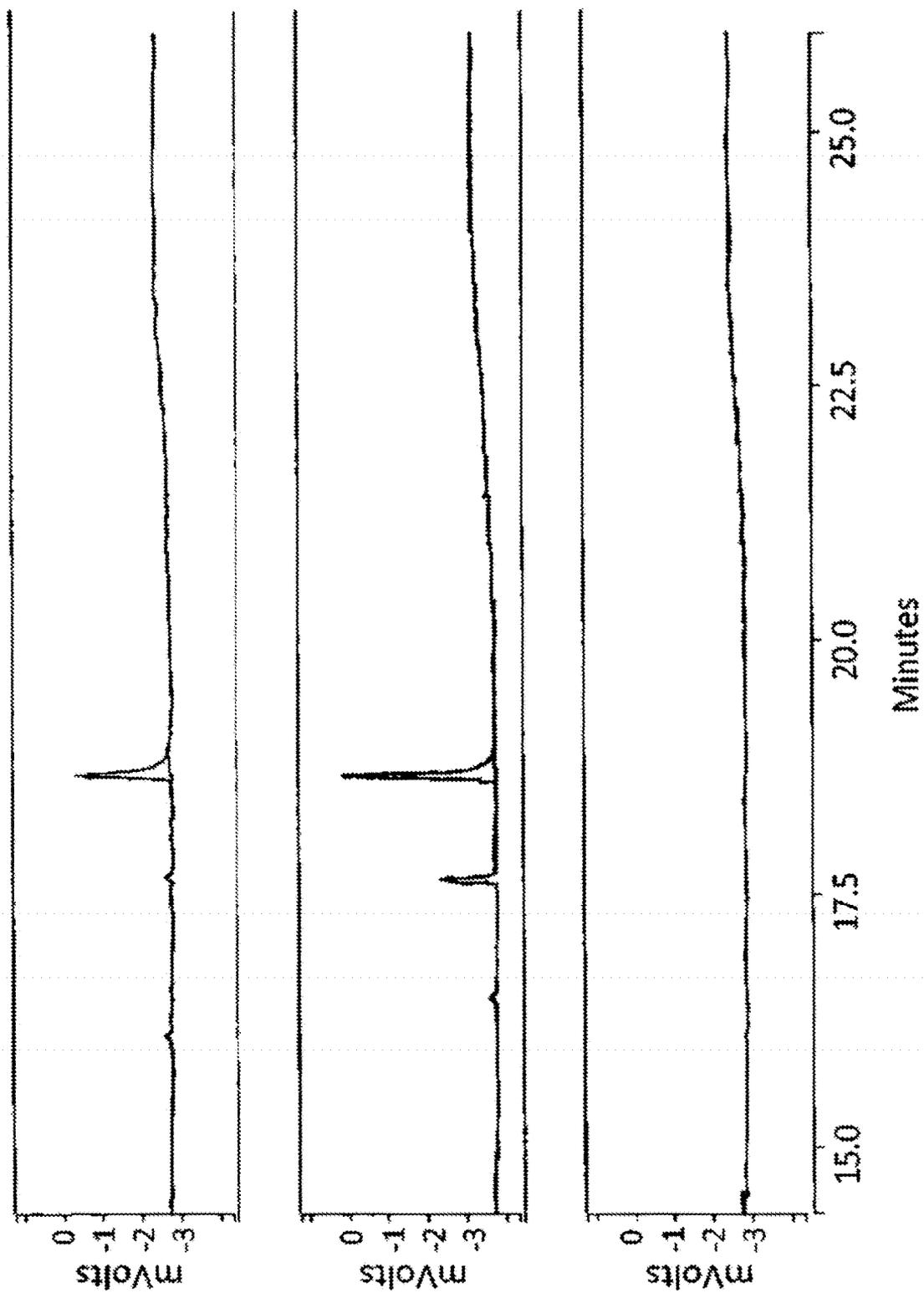
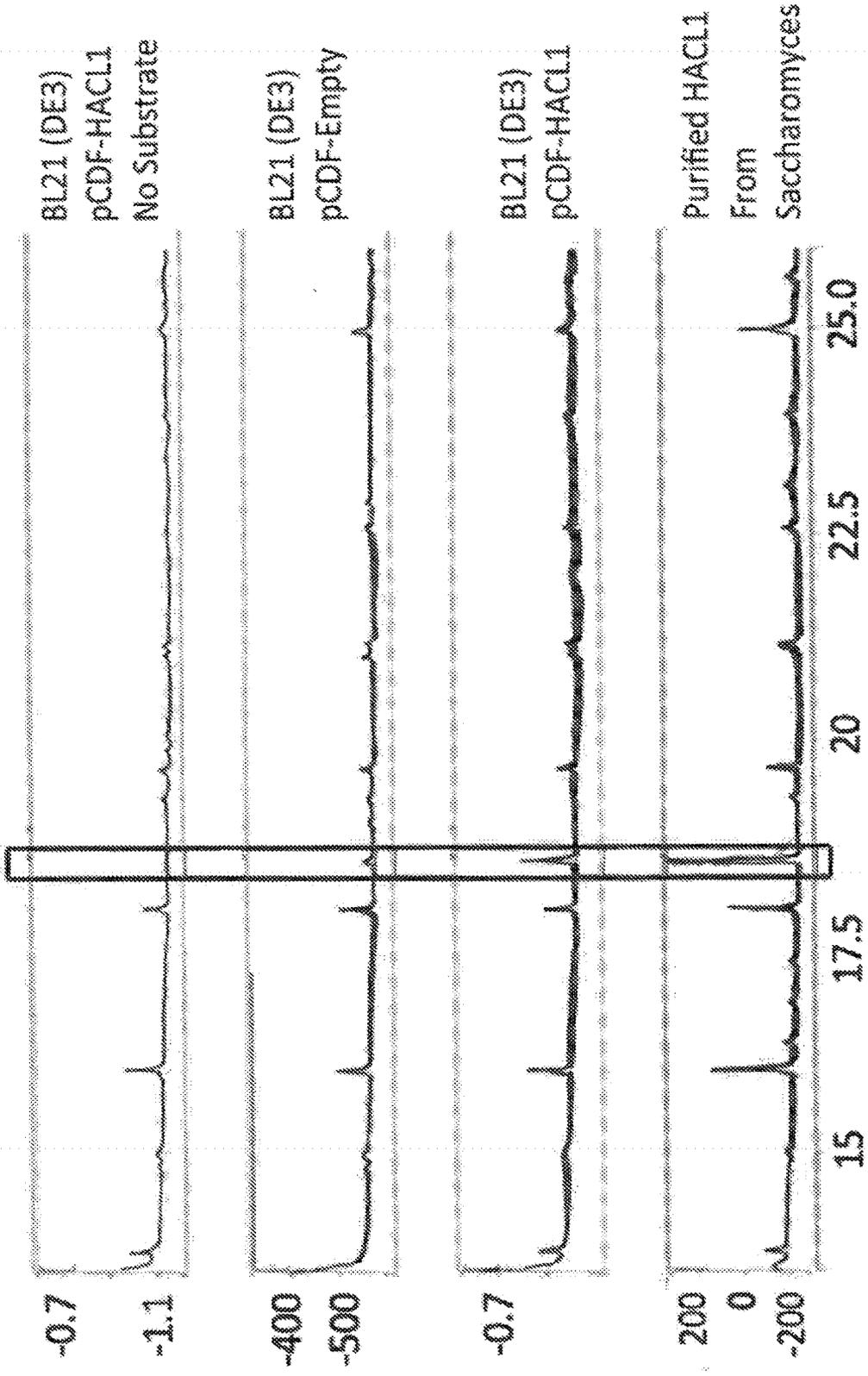
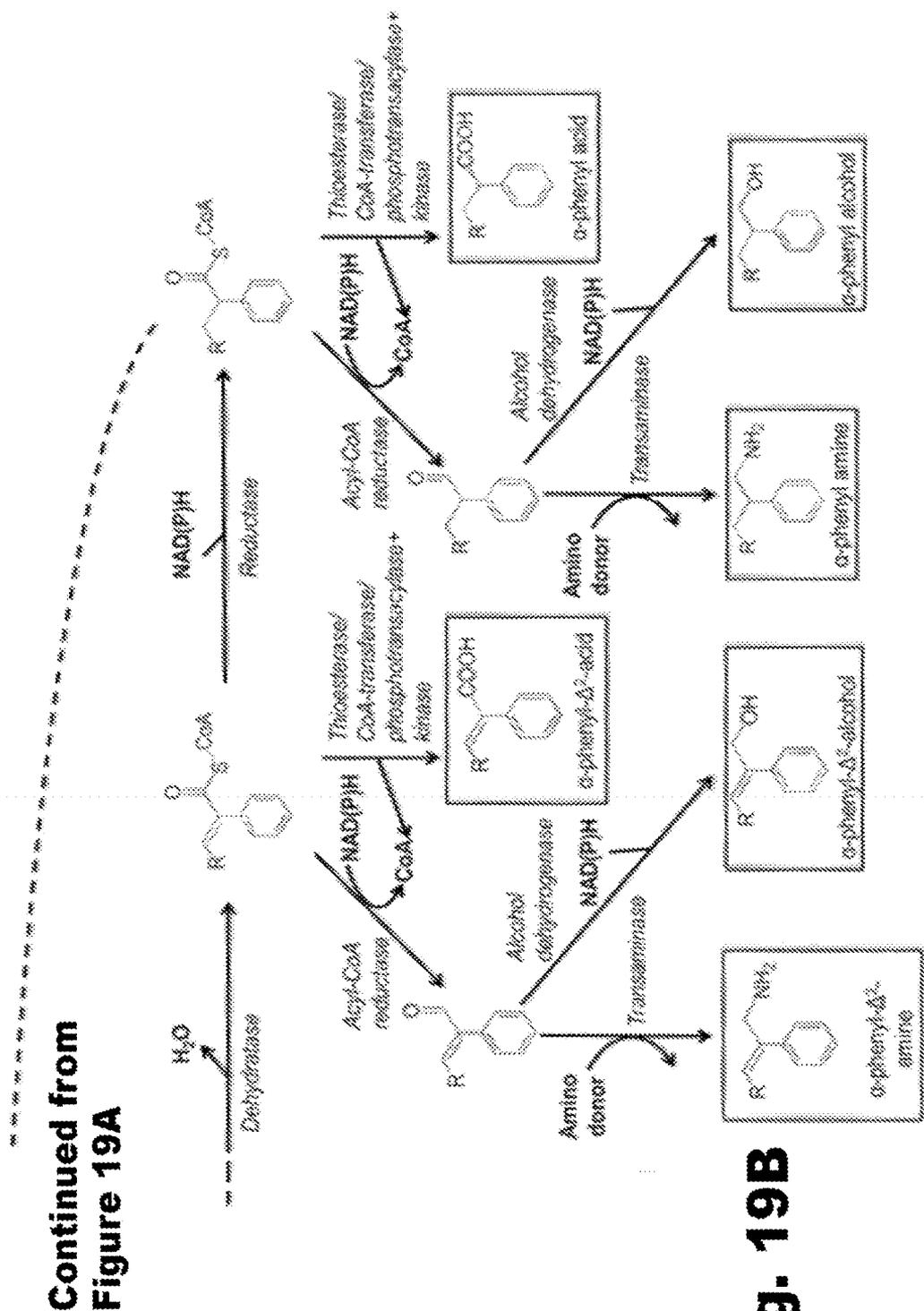


Figure 17



HACL1 expressed with low induction -- 60 minute assay

Figure 18



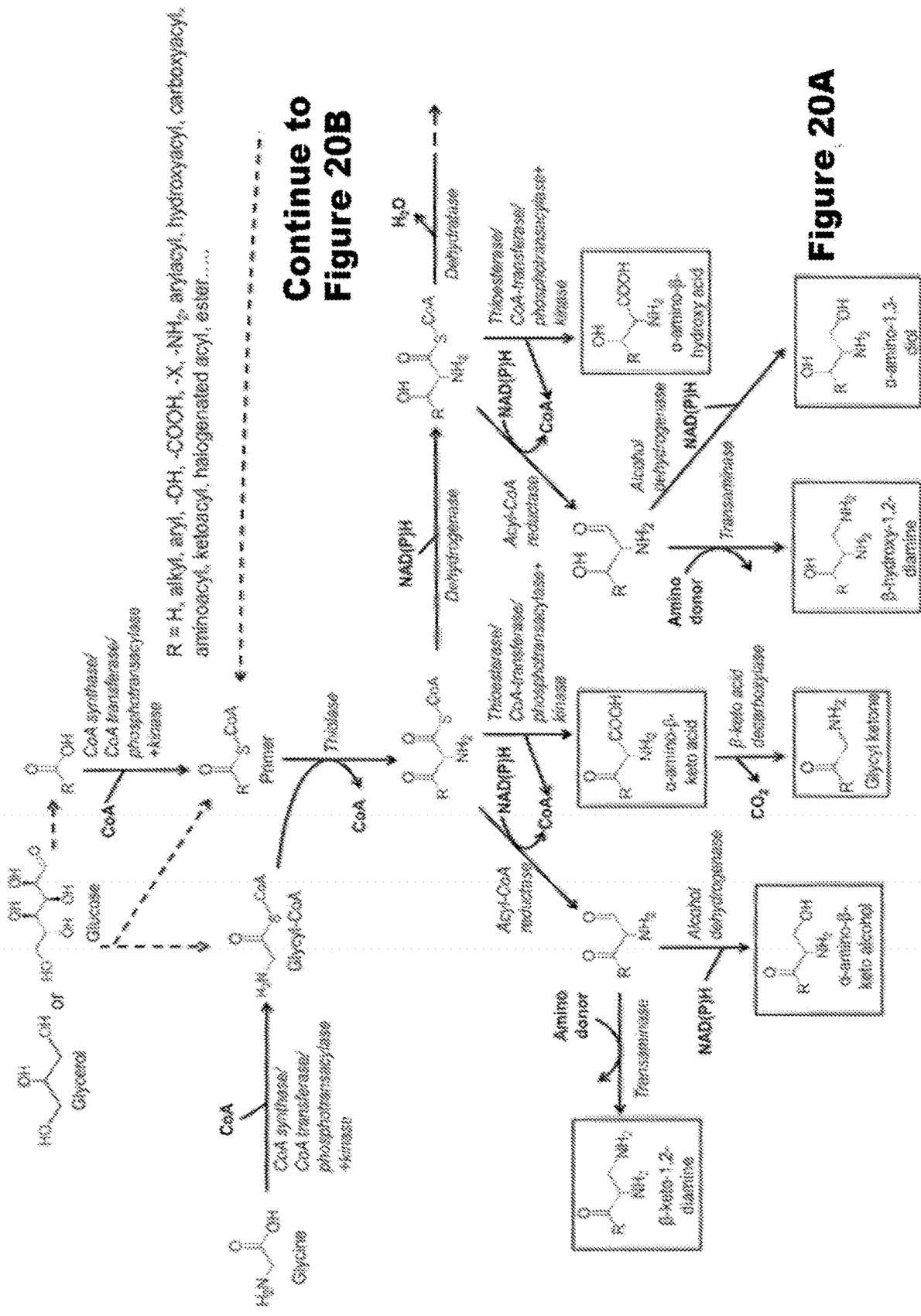


Figure 20A

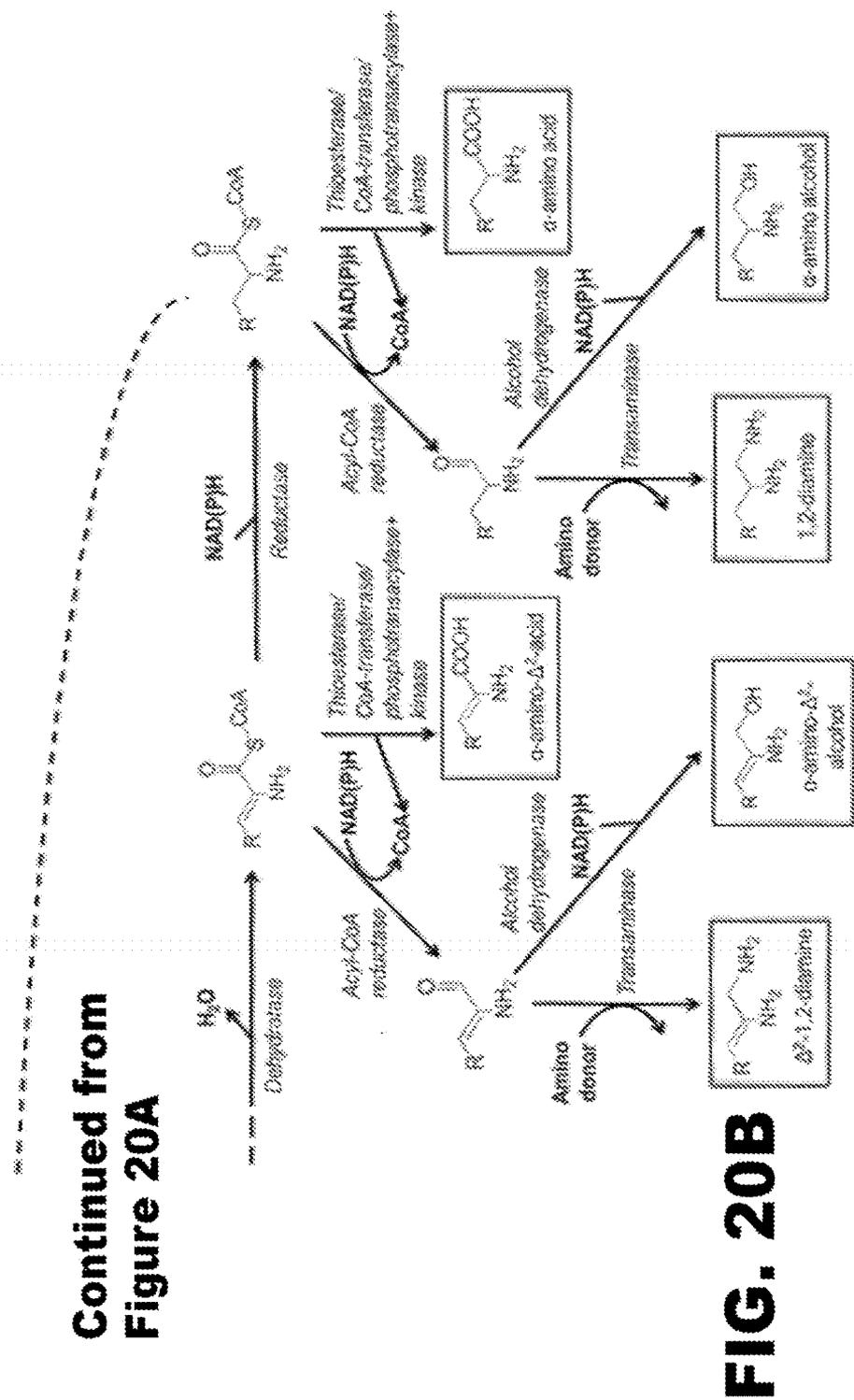


FIGURE 21

<p>A recombinant microorganism comprising overexpressed enzymes including 1) a thiolase catalyzing the condensation of an unsubstituted or functionalized acyl-CoA thioester with alpha-functionalized acetyl-CoA; a 2 hydroxyacyl-CoA dehydrogenase, 3) an enoyl-CoA hydratase, 4) an enoyl-CoA reductase and 5) a termination enzyme such as thioesterase.</p>
<p>A recombinant microorganism comprising an inducible expression vector or inducible integrated sequences for overexpressing enzymes including 1) a thiolase catalyzing the condensation of an unsubstituted or functionalized acyl-CoA thioester with alpha-functionalized acetyl-CoA; a 2 hydroxyacyl-CoA dehydrogenase, 3) an enoyl-CoA hydratase, 4) an enoyl-CoA reductase and 5) a termination enzyme such as thioesterase.</p>
<p>A recombinant microorganism being a bacteria comprising an inducible expression vector or inducible integrated sequences for overexpressing enzymes including 1) a thiolase catalyzing the condensation of an unsubstituted or functionalized acyl-CoA thioester with alpha-functionalized acetyl-CoA; a 2 hydroxyacyl-CoA dehydrogenase, 3) an enoyl-CoA hydratase, 4) an enoyl-CoA reductase and 5) a termination enzyme such as thioesterase.</p>
<p>A recombinant microorganism being a <i>E. coli</i> comprising an inducible expression vector or inducible integrated sequences for overexpressing enzymes including 1) a thiolase catalyzing the condensation of an unsubstituted or functionalized acyl-CoA thioester with alpha-functionalized acetyl-CoA; a 2 hydroxyacyl-CoA dehydrogenase, 3) an enoyl-CoA hydratase, 4) an enoyl-CoA reductase and 5) a termination enzyme such as thioesterase.</p>
<p>A genetically engineered microorganism comprising means for:</p> <ul style="list-style-type: none"> a) an overexpressed activation enzyme(s) able to produce an alpha-functionalized CoA thioester extender unit, wherein said activation enzyme is selected from: <ul style="list-style-type: none"> i) an acyl-CoA synthase which converts the alpha-functionalized CoA thioester extender unit from an alpha-functionalized acid; ii) an acyl-CoA transferase which converts the alpha-functionalized CoA thioester extender unit from an alpha-functionalized acid; iii) a phosphotransacylase and a carboxylate kinase which converts the alpha-functionalized CoA thioester extender unit from an alpha-functionalized acid; iv) other one or more enzymes that allow the production of the alpha-functionalized CoA thioester extender unit from the carbon source without via the alpha-functionalized acid; b) an overexpressed activation enzyme(s) able to produce an acyl-CoA primer wherein said activation enzyme is selected from: <ul style="list-style-type: none"> i) an acyl-CoA synthase which converts the acyl-CoA primer from its acid form; ii) an acyl-CoA transferase which converts the acyl-CoA primer from its acid form; iii) a phosphotransacylase and a carboxylate kinase which converts the acyl-CoA primer from its acid form; iv) other one or more enzymes that allow the production of the acyl-CoA primer from the carbon source without via the alpha-functionalized acid; c) an overexpressed thiolase enzyme that catalyzes the condensation of an acyl-CoA primer with an alpha-functionalized CoA

FIGURE 21 (CONTINUED)

<p>thioester extender unit to form an alpha-functionalized β-ketoacyl-CoA;</p> <p>d) an overexpressed 3-hydroxyacyl-CoA dehydrogenase or 3-oxoacyl-[acyl-carrier-protein] reductase enzyme that catalyzes the reduction of said alpha-functionalized β-ketoacyl-CoA to produce an alpha-functionalized β-hydroxyacyl-CoA;</p> <p>e) an overexpressed enoyl-CoA hydratase, 3-hydroxyacyl-CoA dehydratase, or 3-hydroxyacyl-[acyl-carrier-protein] dehydratase enzyme that catalyzes the dehydration of said alpha-functionalized β-hydroxyacyl-CoA to an alpha-functionalized trans-enoyl-CoA;</p> <p>f) an overexpressed acyl-CoA dehydrogenase, trans-enoyl-CoA reductase, or enoyl-[acyl-carrier-protein] reductase enzyme that catalyzes the reduction of said alpha-functionalized trans-enoyl-CoA to an alpha-functionalized acyl-CoA;</p> <p>g) iterations of steps b to e, wherein said iteration is achieved by utilizing an alpha-functionalized acyl-CoA-thioester product generated in step e of the last turn as an primer or an extender unit of step b in the next turn of iteration;</p> <p>h) an overexpressed termination enzyme(s) able to use a substrate selected from the group consisting of alpha-functionalized β-ketoacyl-CoA-thioester products generated in step b, alpha-functionalized β-hydroxyacyl-CoA-thioester products generated in step c, alpha-functionalized trans-enoyl-CoA-thioester products generated in step d and alpha-functionalized acyl-CoA-thioester products generated in step e, wherein said termination enzyme(s) is selected from:</p> <ol style="list-style-type: none">i) the group consisting of a thioesterase, or an acyl-CoA transferase, or a phosphotransacylase and a carboxylate kinase catalyzing the conversion of the CoA moiety of substrate CoA thioester to a carboxylic acid group;ii) an aldehyde-forming acyl-CoA reductase catalyzing the conversion of the CoA moiety of a substrate to an aldehyde group and an alcohol dehydrogenase catalyzing the conversion of an aldehyde to an alcohol;iii) an aldehyde-forming acyl-CoA reductase catalyzing the conversion of the CoA moiety of a substrate to an aldehyde group and a transaminase catalyzing the conversion of an aldehyde to an amine;i) optionally reduced expressions of fermentation genes leading to reduced production of lactate, acetate, ethanol and succinate; <p>and</p> <p>wherein said microorganism has an iterative carbon elongation pathway beginning with said acyl-CoA thioester primer and alpha-functionalized CoA thioester extender unit and running in a biosynthetic direction.</p>
<p>Any microorganism as herein described, wherein said acyl-CoA primer is an acyl CoA thioester whose omega group is selected from the group consisting of hydrogen, alkyl group, hydroxyl group, carboxyl group, aryl group, halogen, amino group, hydroxyacyl group, carboxyacyl group, aminoacyl group, ketoacyl group, halogenated acyl group, and any other functionalized acyl groups.</p> <p>Any microorganism as herein described, wherein said an alpha-functionalized CoA thioester extender unit is an acyl CoA thioester whose alpha group is selected from the group consisting of hydrogen, alkyl group, hydroxyl group, carboxyl group, aryl group, halogen, amino group, hydroxyacyl group, carboxyacyl group, aminoacyl group, ketoacyl group, halogenated acyl group, and any other functionalized acyl groups.</p>
<p>Any microorganism as herein described, wherein said alpha-functionalized acid is the acid form of alpha-functionalized CoA thioester extender unit whose omega group is selected from the group consisting of hydrogen, alkyl group, hydroxyl group, carboxyl group, aryl</p>

FIGURE 21(CONTINUED)

<p>group, halogen, amino group, hydroxyacyl group, carboxyacyl group, aminoacyl group, ketoacyl group, halogenated acyl group, and any other functionalized acyl groups.</p>
<p>Any microorganism as herein described, wherein said acid form of acyl-CoA primer has omega group selected from the group consisting of hydrogen, alkyl group, hydroxyl group, carboxyl group, aryl group, halogen, amino group, hydroxyacyl group, carboxyacyl group, aminoacyl group, ketoacyl group, halogenated acyl group, and any other functionalized acyl groups.</p>
<p>Any microorganism as herein described, wherein said alpha-functionalized acid is supplemented in the media or supplied through the intracellular pathway from the carbon source.</p>
<p>Any microorganism as herein described, wherein said acid form of acyl-CoA primer is supplemented in the media or supplied through the intracellular pathway from the carbon source.</p>
<p>Any microorganism as herein described, wherein said genetically engineered microorganism produces a product selected from the group consisting of β-keto acids, β-keto alcohols, β-keto amines, β-hydroxy acids, 1,3-diols, β-hydroxy amines, Δ^2-fatty alcohols, Δ^2-amines, fatty acids, alcohols and amines whose alpha group is selected from the group consisting of hydrogen, alkyl group, hydroxyl group, carboxyl group, aryl group, halogen, amino group, hydroxyacyl group, carboxyacyl group, aminoacyl group, ketoacyl group, halogenated acyl group, and any other functionalized acyl groups.</p>
<p>Any microorganism as herein described, wherein said step g uses alpha-functionalized β-ketoacyl-CoA-thioester products generated in step b as the substrate, further comprising an overexpressed β-keto acid decarboxylase catalyzing the conversion of the β-keto-acid to a ketone.</p>
<p>Any microorganism as herein described, wherein said genetically engineered microorganism produces a ketone whose omega group is selected from the group consisting of hydrogen, alkyl group, hydroxyl group, carboxyl group, aryl group, halogen, amino group, hydroxyacyl group, carboxyacyl group, aminoacyl group, ketoacyl group, halogenated acyl group, and any other functionalized acyl groups.</p>
<p>Any microorganism as herein described, wherein said termination pathway i) of step g uses alpha-functionalized acyl-CoA-thioester products generated in step b as the substrate, utilizing glycoyl-CoA as the extender unit and further comprising: an overexpressed keto-dehydrogenase catalyzing the conversion of a 2-hydroxy acid to an alpha-keto acid; an overexpressed alpha-keto acid decarboxylase catalyzing the conversion of an alpha-keto acid to a primary aldehyde; an overexpressed alcohol dehydrogenase catalyzing the conversion of a primary aldehyde to a primary alcohol.</p>
<p>Any microorganism as herein described, utilizing glycoyl-CoA as the extender unit and further comprising: an overexpressed 2-hydroxyacyl-CoA lyase catalyzing the conversion of a 2-hydroxyacyl-CoA, generated from step e, to a primary aldehyde and a formyl-CoA; an overexpressed alcohol dehydrogenase catalyzing the conversion of a primary aldehyde to a primary alcohol.</p>
<p>Any microorganism as herein described, wherein said genetically engineered microorganism produces a primary alcohol.</p>
<p>Any microorganism as herein described, wherein said overexpressed acyl-CoA synthase is encoded by a gene(s) selected from the group consisting of <i>E. coli sucC</i>, <i>E. coli sucD</i>, <i>E. coli paaK</i>, <i>E. coli prpE</i>, <i>E. coli menE</i>, <i>E. coli fadK</i>, <i>E. coli fadD</i>, <i>Penicillium</i></p>

FIGURE 21 (CONTINUED)

<p><i>chrysogenum phi</i>, <i>Salmonella typhimurium</i> LT2 prpE, <i>Bacillus subtilis</i> bioW, <i>Cupriavidus basilensis</i> hmfD, <i>Rhodospseudomonas palustris</i> badA, <i>R. palustris</i> hbaA, <i>Pseudomonas aeruginosa</i> PAO1 pqsA, <i>Arabidopsis thaliana</i> 4cl and other homologs.</p> <p>Any microorganism as herein described, wherein said overexpressed acyl-CoA transferase is encoded by a gene(s) selected from the group consisting of <i>E. coli</i> atoD, <i>E. coli</i> sopC, <i>E. coli</i> ydiF, <i>E. coli</i> atoA, <i>E. coli</i> atoD, <i>Clostridium acetobutylicum</i> ctfA, <i>C. acetobutylicum</i> ctfB, <i>Clostridium kluuyveri</i> cat2, <i>C. kluuyveri</i> cat1, <i>P. putida</i> pcaI, <i>P. putida</i> pcaJ, <i>Megasphaera elsdenii</i> pct, <i>Acidaminococcus fermentans</i> gctA, <i>Acidaminococcus fermentans</i> gctB, <i>Acetobacter acetii</i> aarC and other homologs.</p> <p>Any microorganism as herein described, wherein said overexpressed phosphotransacylase is encoded by a gene(s) selected from the group consisting of <i>Clostridium acetobutylicum</i> ptb, <i>Enterococcus faecalis</i> ptb, <i>Salmonella enterica</i> pduL and other homologs.</p> <p>Any microorganism as herein described, wherein said overexpressed carboxylate kinase is encoded by a gene(s) selected from the group consisting of <i>Clostridium acetobutylicum</i> buk, <i>Enterococcus faecalis</i> buk, <i>Salmonella enterica</i> pduW and other homologs.</p> <p>Any microorganism as herein described, wherein said overexpressed thiolase is encoded by a gene(s) selected from the group consisting of <i>E. coli</i> atoB, <i>E. coli</i> yqeF, <i>E. coli</i> fadA, <i>E. coli</i> fadI, <i>Ralstonia eutropha</i> bktB, <i>Pseudomonas</i> sp. B13 catF, <i>E. coli</i> paaJ, <i>Rhodococcus opacus</i> pcaF, <i>Pseudomonas putida</i> pcaF, <i>Streptomyces</i> sp. pcaF, <i>P. putida</i> fadAx, <i>P. putida</i> fadA, <i>Ralstonia eutropha</i> phaA, <i>Acinetobacter</i> sp. ADP1 dcaF, <i>Clostridium acetobutylicum</i> thIA, <i>Clostridium acetobutylicum</i> thIB and other homologs.</p> <p>Any microorganism as herein described, wherein said overexpressed 3-hydroxyacyl-CoA dehydrogenase or 3-oxoacyl-[acyl-carrier-protein] reductase is encoded by a gene(s) selected from the group consisting of <i>E. coli</i> fabG, <i>E. coli</i> fadB, <i>E. coli</i> fadJ, <i>E. coli</i> paaH, <i>P. putida</i> fadB, <i>P. putida</i> fadB2x, <i>Acinetobacter</i> sp. ADP1 dcaH, <i>Ralstonia eutropha</i> phaB, <i>Clostridium acetobutylicum</i> hbd and other homologs.</p> <p>Any microorganism as herein described, wherein said overexpressed enoyl-CoA hydratase, 3-hydroxyacyl-CoA dehydratase, or 3-hydroxyacyl-[acyl-carrier-protein] dehydratase is encoded by a gene(s) selected from the group consisting of <i>E. coli</i> fabA, <i>E. coli</i> fabZ, <i>E. coli</i> fadB, <i>E. coli</i> fadJ, <i>E. coli</i> paaF, <i>P. putida</i> fadB, <i>P. putida</i> fadB1x, <i>Acinetobacter</i> sp. ADP1 dcaE, <i>Clostridium acetobutylicum</i> crt, <i>Aeromonas caviae</i> phaJ and other homologs.</p> <p>Any microorganism as herein described, wherein said acyl-CoA dehydrogenase, trans-enoyl-CoA reductase, or enoyl-[acyl-carrier-protein] reductase is encoded by a gene(s) selected from the group consisting of <i>E. coli</i> fadE, <i>E. coli</i> ydiO, <i>Euglena gracilis</i> TER, <i>Treponema denificola</i> TER, <i>Clostridium acetobutylicum</i> TER, <i>E. coli</i> fabI, <i>Enterococcus faecalis</i> fabK, <i>Bacillus subtilis</i> fabL, <i>Vibrio cholerae</i> fabV and other homologs.</p> <p>Any microorganism as herein described, wherein said overexpressed thioesterase is encoded by a gene(s) selected from the group consisting of <i>E. coli</i> tesA, <i>E. coli</i> tesB, <i>E. coli</i> yciA, <i>E. coli</i> fadM, <i>E. coli</i> ydiI, <i>E. coli</i> ybgC, <i>E. coli</i> paaI, <i>Mus musculus</i> acof8, <i>Alcanivorax borkumensis</i> tesB2, <i>Fibrobacter succinogenes</i> Fs2108, <i>Prevotella ruminicola</i> Pr655, <i>Prevotella ruminicola</i> Pr1687, <i>Lycopersicon hirsutum</i> 1 glabratum mks2 and other homologs.</p> <p>Any microorganism as herein described, wherein said overexpressed aldehyde-forming acyl-CoA reductase is encoded by a gene(s) selected from the group consisting <i>Acinetobacter calcoaceticus</i> acr1, <i>Acinetobacter</i> sp Strain M-1 acrM, <i>Clostridium beijerinckii</i> ald, <i>E. coli</i> eufE, <i>Salmonella enterica</i> eufE, <i>E. coli</i> mhpF, <i>Clostridium kluuyveri</i> sucD and other homologs.</p>

FIGURE 21 (CONTINUED)

<p>Any microorganism as herein described, wherein said overexpressed alcohol dehydrogenase is encoded by a gene(s) selected from the group consisting <i>E. coli betaA</i>, <i>E. coli dkgA</i>, <i>E. coli eufG</i>, <i>E. coli fucO</i>, <i>E. coli ucpA</i>, <i>E. coli yahK</i>, <i>E. coli ybbO</i>, <i>E. coli ybdH</i>, <i>E. coli yiaY</i>, <i>E. coli yigB</i>, <i>Saccharomyces cerevisiae ADH6</i>, <i>Clostridium kluyveri 4hbD</i>, <i>Acinetobacter</i> sp. SE19 <i>chnD</i> and other homologs.</p> <p>Any microorganism as herein described, wherein said overexpressed transaminase is encoded by a gene(s) selected from the group consisting of <i>Arabidopsis thaliana</i> At3g22200, <i>Alcaligenes denitrificans aptA</i>, <i>Bordetella bronchiseptica</i> BB0869, <i>Bordetella parapertussis</i> BPP0784, <i>Brucella melitensis</i> BAWG_0478, <i>Burkholderia pseudomallei</i> BP1026B_10669, <i>Chromobacterium violaceum</i> CV2025, <i>Oceanicola granulosus</i> OG2516_07293, <i>Paracoccus denitrificans</i> PD1222 Pden_3984, <i>Caulobacter crescentus</i> CC_3143, <i>Pseudogulbenkiania ferrooxidans</i> w-TA, <i>Pseudomonas putida</i> w -TA, <i>Ralstonia solanacearum</i> w -TA, <i>Rhizobium meliloti</i> SMC01534, <i>Vibrio fluvialis</i> w -TA, <i>Bacillus megaterium</i> SC6394 w -TA, <i>Mus musculus abaT</i>, <i>Flavobacterium lutescens</i> lat, <i>Streptomyces clavuligerus</i> lat, <i>E. coli gabT</i>, <i>E. coli puuE</i>, <i>E. coli ygjG</i> and other homologs.</p> <p>Any microorganism as herein described, wherein said overexpressed β-keto acid decarboxylase is encoded by a gene(s) selected from the group consisting of <i>Clostridium acetobutylicum</i> adc, <i>Lycopersicon hirsutum</i> f <i>glabratum mks1</i> and other homologs.</p> <p>Any microorganism as herein described, wherein said overexpressed keto-dehydrogenase is encoded by a gene(s) selected from the group consisting of <i>E. coli ldhA</i>, <i>E. coli lldD</i>, <i>E. coli leuB</i>, <i>Clostridium beijerinckii adh</i>, <i>Acidaminococcus fermentans hgdH</i>, <i>E. coli serA</i>, <i>Gordonia</i> sp. TY-5 <i>adh1</i>, <i>Gordonia</i> sp. TY-5 <i>adh2</i>, <i>Gordonia</i> sp. TY-5 <i>adh3</i>, <i>Rhodococcus ruber adh-A</i> and other homologs.</p> <p>Any microorganism as herein described, wherein said overexpressed alpha-keto acid decarboxylase is encoded by a gene(s) selected from the group consisting <i>Lactococcus lactis kivd</i>, <i>Saccharomyces cerevisiae PDC1</i>, <i>S. cerevisiae PDC5</i>, <i>S. cerevisiae PDC6</i>, <i>S. cerevisiae ARO10</i>, <i>S. cerevisiae THI3</i>, <i>Zymomonas mobilis pdc</i> and other homologs.</p> <p>Any microorganism as herein described, wherein said overexpressed 2-hydroxyacyl-CoA lyase is encoded by a gene(s) selected from the group consisting <i>Homo sapiens hac11</i>, <i>Rattus norvegicus hac11</i>, <i>Dictyostelium discoideum hac11</i>, <i>Mus musculus hac11</i> and other homologs.</p> <p>Any microorganism as herein described, wherein said reduced expressions of fermentation enzymes are $\Delta adhE$, (Δpta or $\Delta ackA$ or $\Delta ackApta$), $\Delta poxB$, $\Delta ldhA$, and $\Delta frdA$ and less acetate, lactate, ethanol and succinate are thereby produced.</p> <p>Any microorganism as herein described, comprising one or more of the following mutations: <i>fadR</i>, <i>atoC(c)</i>, $\Delta arcA$, Δcrp, <i>crp*</i>.</p> <p>Any microorganism herein described, said overexpressed enzymes being under the control of an inducible promoter, preferably multiple enzymes under the control of a single promoter, preferably allowing for coordinate expression of the enzymes. In one embodiment, the enzymes are expressed off an expression vector, but in another one or more are integrated into the genome.</p> <p>A method of making alpha functionalized products, comprising growing any microorganism described herein in a nutrient broth under conditions such that said enzymes are overexpressed, said microorganism producing alpha functionalized product using said overexpressed enzymes, and isolating said alpha functionalized product.</p>

ITERATIVE PLATFORM FOR THE SYNTHESIS OF ALPHA FUNCTIONALIZED PRODUCTS

PRIOR RELATED APPLICATIONS

[0001] This application is a continuation-in-part of U.S. Ser. No. 16/818,642 filed on Mar. 13, 2020, which is a continuation-in-part of U.S. Ser. No. 15/566,704 filed on Oct. 14, 2017 (now abandoned), which is a National Phase under 35 U.S.C. § 371 of International Application PCT/US2016/027873, filed Apr. 15, 2016, which claims priority to U.S. Ser. No. 62/148,123, ITERATIVE PLATFORM FOR THE SYNTHESIS OF ALPHA FUNCTIONALIZED PRODUCTS, filed Apr. 15, 2015. All applications are expressly incorporated by reference herein in their entirety for all purposes.

FEDERALLY SPONSORED RESEARCH STATEMENT

[0002] This invention was made with government support under Grant Nos: CBET1067565 and CBET1134541 awarded by the National Science Foundation. The government has certain rights in the invention.

REFERENCE TO ELECTRONIC SEQUENCE LISTING

[0003] This application contains a Sequence Listing which has been submitted electronically in .XML format and is hereby incorporated in its entirety. Said .XML copy, created on Jul. 1, 2023, is named "GON2015-080-CIP2.xml" and is 26,469 bytes in size. The sequence listing contained in this .XML file is part of the specification and is hereby incorporated by reference herein in its entirety.

FIELD OF THE DISCLOSURE

[0004] This disclosure generally relates to the use of recombinant microorganisms to make various products.

BACKGROUND OF THE DISCLOSURE

[0005] Reactions that catalyze the iterative formation of carbon-carbon bonds are instrumental for many metabolic pathways, such as the biosynthesis of fatty acids, polyketides, and many other molecules with applications ranging from biofuels and green chemicals to therapeutic agents. These pathways typically start with small precursor metabolites that serve as building blocks that are subsequently condensed and modified in an iterative fashion until the desired chain length and functionality are achieved.

[0006] Most iterative carbon-carbon bond forming reactions in natural biological systems take place through a Claisen condensation mechanism in which the nucleophilic α -anion of an acyl-thioester, serving as the extender unit, attacks the electrophilic carbonyl carbon of another acyl-thioester, serving as the primer. Depending on how the nucleophilic α -anion is generated, the Claisen condensation reaction can be classified as decarboxylative or non-decarboxylative.

[0007] Many natural iterative carbon chain elongation pathways, like fatty acid and polyketide biosynthesis pathways, utilize decarboxylative Claisen condensation reactions with malonyl thioesters as extender units. Their potential products include fatty acids, alcohols, polyketides,

esters, alkanes and alkenes with diverse chain lengths, structures and functionalities due to usage of functionalized primers, usage of α -functionalized malonyl thioesters as extender units and diverse pathways for termination of carbon chain elongation and subsequent product modification. However, despite the structural and functional diversity of these products, the use of malonyl thioester as a C2 extender unit requires the ATP-dependent activation of acetyl-CoA to malonyl-CoA, which in turn limits the energy efficiency of these pathways. Furthermore, owing to the decarboxylation mechanism, the β -site of extender units of the decarboxylative Claisen condensation must be a carboxylic group, restricting the range of extender units and potentially limiting the diversity of products that can be generated through these carbon chain elongation pathways.

[0008] In order to overcome this limitation, we have recently implemented a novel approach by driving beta-oxidation in reverse to make fatty acids instead of degrading them (see U.S. 20130316413, WO2013036812, each incorporated by reference in its entirety for all purposes). Unlike the fatty acid biosynthesis pathway, the reversal of the β -oxidation cycle operates with coenzyme-A (CoA) thioester intermediates and uses acetyl-CoA directly for acyl-chain elongation (rather than first requiring ATP-dependent activation to malonyl-CoA). In these pathways, thiolases catalyze the non-decarboxylative Claisen condensation in which acetyl-CoA, instead of malonyl thioesters, serves as the extender unit, and subsequent β -reduction reactions by hydroxyacyl-CoA dehydrogenases (HACDs), enoyl-CoA hydratases (ECHs) and enoyl-CoA reductases (ECRs) enable iteration. Compared to pathways utilizing decarboxylative Claisen condensation, these pathways are more energy efficient due to less ATP consumption for the supply of extender unit acetyl-CoA than malonyl thioesters. However, these thiolases only utilize acetyl-CoA as the extender unit, thus limiting the functionality of synthesized products. A novel non-decarboxylative Claisen condensation reaction able to accept wider range of extender units and proceed in an iterative manner is required to diversify the product range of carbon-chain elongation.

[0009] This disclosure demonstrates a general CoA-dependent carbon elongation platform based on the use of de novo thiolase-catalyzed non-decarboxylative Claisen condensation which accepts functionalized primers and extender units, along with suitable HACDs, ECHs and ECRs (FIG. 1A and FIG. 1B) to complete one turn of the 2-carbon additive cycle. Wide-ranging product diversity (FIG. 1A and FIG. 1B) from this iterative platform is achieved through the use of primers with or without functionalization (R1 in FIG. 1A and FIG. 1B) and extender units with alpha-functionalization (R2 in FIG. 1A and FIG. 1B) in combination with pathway termination to various product classes by multiple pathways from any intermediate with various β -reduction degrees. The proposed platform possesses the potential for the high product diversity of a biosynthetic pathway combined with the high efficiency of a fermentative pathway.

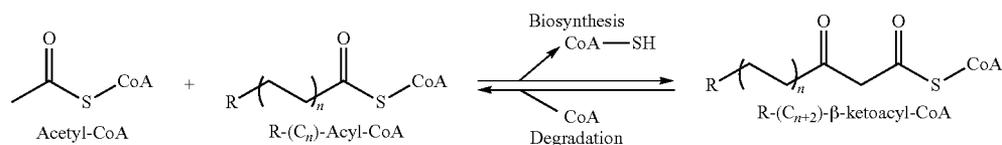
SUMMARY OF THE DISCLOSURE

[0010] This disclosure generally relates to the use of microorganisms to make alpha-functionalized chemicals and fuels, (e.g. alpha-functionalized carboxylic acids, alcohols, hydrocarbons, amines, and their beta-, and omega-functionalized derivatives), by utilizing an iterative carbon chain elongation pathway that uses functionalized extender

units. The core enzymes in the pathway include thiolases, dehydrogenases, dehydratases and reductases. Native or engineered thiolases catalyze the condensation of either unsubstituted or functionalized acyl-CoA primers with an alpha-functionalized acetyl-CoA as the extender unit to generate alpha-functionalized β -keto acyl-CoA. Dehydrogenases convert alpha-functionalized β -keto acyl-CoA to alpha-functionalized β -hydroxy acyl-CoA. Dehydratases convert alpha-functionalized β -hydroxy acyl-CoA to alpha-functionalized enoyl-CoA. Reductases convert alpha-func-

[0015] As used herein, an “extender unit” is the donor of carbons in each cycle of the iterative carbon elongation platform. In this disclosure, the extender unit is alpha-functionalized acetyl-CoAs.

[0016] Thiolases are ubiquitous enzymes that have key roles in many vital biochemical pathways, including the beta-oxidation pathway of fatty acid degradation and various biosynthetic pathways. Members of the thiolase family can be divided into two broad categories: degradative thiolases (EC 2.3.1.16), and biosynthetic thiolases (EC 2.3.1.9). The forward and reverse reactions are shown below:



tionized enoyl-CoA to alpha-functionalized acyl-CoA. The platform can be operated in an iterative manner (i.e. multiple turns) by using the resulting alpha-functionalized acyl-CoA as primer and either acetyl-CoA or the aforementioned alpha-functionalized extender unit in subsequent turns of the cycle. Termination pathways acting on any of the four alpha-functionalized CoA thioester intermediates terminate the platform and generate various alpha-functionalized carboxylic acids, alcohols and amines with different β -reduction degrees.

[0011] This disclosure demonstrates a general CoA-dependent carbon elongation platform based on the use of thiolase-catalyzed non-decarboxylative Claisen condensations that accept alpha-functionalized extender units, along with suitable hydroxyacyl-CoA dehydrogenases (HACDs), enoyl-CoA hydratases (ECHs) and enoyl-CoA reductases (ECRs). A wide-range of alpha-functionalized product families (e.g. alpha-functionalized carboxylic acids, alcohols, hydrocarbons, amines, and their beta-, and omega-functionalized derivatives) can be obtained through this iterative platform.

[0012] The technology entails developing a new pathway that is based on native or engineered thiolases capable of catalyzing the condensation of either unsubstituted or functionalized acyl-CoA primers with an alpha-functionalized acetyl-CoA as the extender unit. This has been reported in neither the scientific, peer-reviewed literature nor the patent literature.

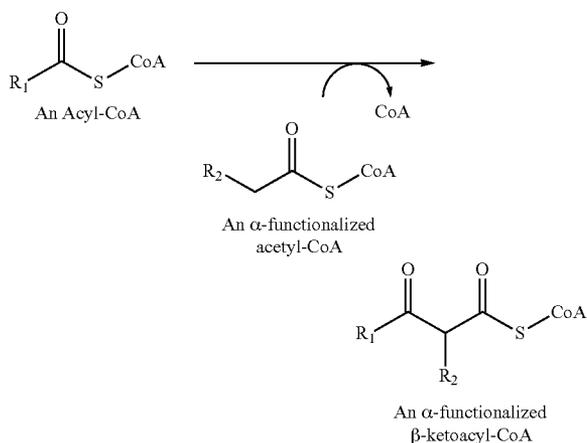
[0013] The process involves performing traditional fermentations using industrial organisms (such as *E. coli*, *S. cerevisiae*) that convert different feedstocks into longer-chain products (e.g. alpha-functionalized carboxylic acids, alcohols, amines, and their beta-, and omega-functionalized derivatives or hydrocarbons). These organisms are considered workhorses of modern biotechnology. Media preparation, sterilization, inoculum preparation, and fermentation are the main steps of the process.

[0014] As used herein, a “primer” is a starting molecule for iterative carbon elongation platform. The “initial primer” or “initiating primer” can be simply acetyl-CoA or other unsubstituted or functionalized acyl-CoAs. As the chain grows by adding extender units in each cycle, the primer will accordingly increase in size.

[0017] These two different types of thiolase are found both in eukaryotes and in prokaryotes: acetoacetyl-CoA thiolase (EC:2.3.1.9) and 3-ketoacyl-CoA thiolase (EC:2.3.1.16). 3-ketoacyl-CoA thiolase (also called thiolase I) has a broad chain-length specificity for its substrates and is involved in degradative pathways such as fatty acid beta-oxidation. Acetoacetyl-CoA thiolase (also called thiolase II) is specific for the thiolysis of acetoacetyl-CoA and involved in biosynthetic pathways such as poly beta-hydroxybutyric acid synthesis or steroid biogenesis.

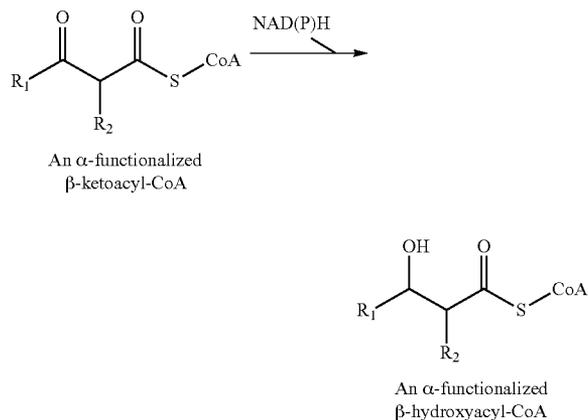
[0018] Furthermore, the degradative thiolases can be made to run in the forward direction by building up the level of left hand side reactants (primer and extender unit), thus driving the equilibrium in the forward direction and/or by overexpressing same or by expressing a mutant of same.

[0019] As used herein, a “thiolase” is an enzyme that catalyzes the condensation of an unsubstituted or functionalized acyl-CoA thioester with alpha-functionalized acetyl-CoA as the carbon donor for chain elongation to produce an unsubstituted or omega-functionalized alpha-functionalized β -keto acyl-CoA in a non-decarboxylative condensation reaction:

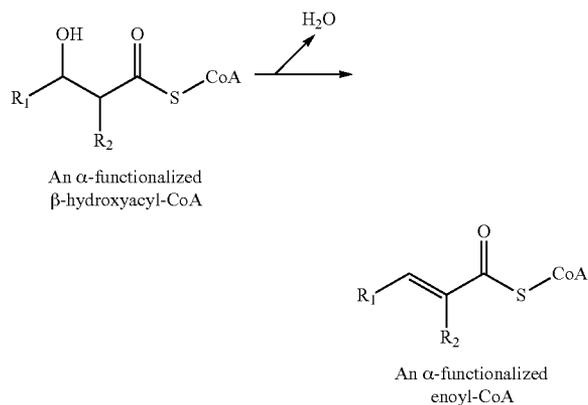


[0020] As used herein, a “hydroxyacyl-CoA dehydrogenase” or “HACD”, is an enzyme that catalyzes the reduction of an unsubstituted or omega-functionalized alpha-func-

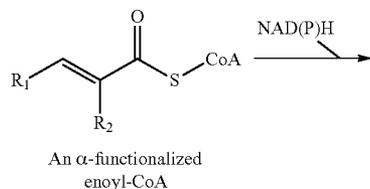
alized β -keto acyl-CoA to an unsubstituted or omega-functionalized alpha-functionalized β -hydroxy acyl-CoA:



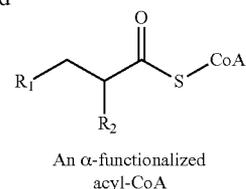
[0021] As used herein, an “enoyl-CoA hydratase” or “ECH” is an enzyme that catalyzes the dehydration of an unsubstituted or omega-functionalized or alpha-functionalized β -hydroxy acyl-CoA to an unsubstituted or omega-functionalized or alpha-functionalized enoyl-CoA:



[0022] As used herein, an “enoyl-CoA reductase” or “ECR” is an enzyme that catalyzes the reduction of an unsubstituted or omega-functionalized or alpha-functionalized transenoyl-CoA to an unsubstituted or omega-functionalized or alpha-functionalized acyl-CoA:



-continued



[0023] As used herein, “termination pathway” refers to one or more enzymes (or genes encoding same) that will pull reaction CoA thioester intermediates out the iterative cycle and produce the desired end product.

[0024] As used herein, an “alpha functionalized product” is a carboxylic acid, alcohols, hydrocarbons, or amine, wherein the alpha position is the second carbon and has an R group that is not hydrogen (R preferably being e.g., alkyl, aryl, —OH, —COOH, or —X, but including others). Note that the second carbon is defined with respect to the -CoA end, and the numbering is retained even when the -CoA is removed. Such alpha functionalized products can be further modified herein, and thus include beta-, and omega-functionalized derivatives.

[0025] As used herein, the expressions “microorganism,” “microbe,” “strain” and the like may be used interchangeably and all such designations include their progeny. It is also understood that all progeny may not be precisely identical in DNA content, due to deliberate or inadvertent mutations. Mutant progeny that have the same function or biological activity as screened for in the originally transformed cell are included. Where distinct designations are intended, it will be clear from the context.

[0026] As used herein, reference to a “cell” is generally understood to include a culture of such cells, as the work described herein is done in cultures having 10^9 - 10^{15} cells.

[0027] As used herein, “growing” cells used in its art accepted manner, referring to exponential growth of a culture of cells, not the few cells that may not have completed their cell cycle at stationary phase or have not yet died in the death phase or after harvesting.

[0028] As used in the claims, “homolog” means an enzyme with at least 50% identity to one of the listed sequences and also having the same general catalytic activity, although of course Km, Kcat and the like can vary. While higher identity (60%, 70%, 80%) and the like may be preferred, it is typical for bacterial sequences to diverge significantly (40-60%), yet still be identifiable as homologs, while mammalian species tend to diverge less (80-90%).

[0029] Reference to proteins herein can be understood to include reference to the gene encoding such protein. Thus, a claimed “permease” protein can include the related gene encoding that permease. However, it is preferred herein to refer to the protein by standard name per ecoliwiki or HUGO since both enzymatic and gene names have varied widely, especially in the prokaryotic arts.

[0030] Once an exemplary protein is obtained, many additional examples of proteins with similar activity can be identified by BLAST search. Further, every protein record is linked to a gene record, making it easy to design overexpression vectors. Many of the needed enzymes are already available in vectors, and can often be obtained from cell depositories or from the researchers who cloned them. But, if necessary, new clones can be prepared based on available

sequence information using RT-PCR techniques. Thus, it should be easily possible to obtain all of the needed enzymes for overexpression.

[0031] Another way of finding suitable enzymes/proteins for use in the invention is to consider other enzymes with the same EC number, since these numbers are assigned based on the reactions performed by a given enzyme. An enzyme that thus be obtained, e.g., from AddGene or from the author of the work describing that enzyme, and tested for functionality as described herein. In addition, many sites provide lists of proteins that all catalyze the same reaction.

[0032] Understanding the inherent degeneracy of the genetic code allows one of ordinary skill in the art to design multiple nucleotides that encode the same amino acid sequence. NCBI™ provides codon usage databases for optimizing DNA sequences for protein expression in various species. Using such databases, a gene or cDNA may be “optimized” for expression in *E. coli*, yeast, algal or other species using the codon bias for the species in which the gene will be expressed.

[0033] Initial cloning experiments have proceeded in *E. coli* for convenience since most of the required genes are already available in plasmids suitable for bacterial expression, but the addition of genes to bacteria is of nearly universal applicability. Indeed, since recombinant methods were invented in the 70's and are now so commonplace, even school children perform genetic engineering experiments using bacteria. Such species include e.g., *Bacillus*, *Streptomyces*, *Azotobacter*, *Trichoderma*, *Rhizobium*, *Pseudomonas*, *Micrococcus*, *Nitrobacter*, *Proteus*, *Lactobacillus*, *Pediococcus*, *Lactococcus*, *Salmonella*, *Streptococcus*, *Paracoccus*, *Methanosarcina*, and *Methylococcus*, or any of the completely sequenced bacterial species. Indeed, hundreds of bacterial genomes have been completely sequenced, and this information greatly simplifies both the generation of vectors encoding the needed genes, as well as the planning of a recombinant engineering protocol. Such species are listed along with links at en.wikipedia.org/wiki/List_of_sequenced_bacterial_genomes.

[0034] Additionally, yeasts, such as *Saccharomyces*, are a common species used for microbial manufacturing, and many species can be successfully transformed. Indeed, yeast are already available that express recombinant thioesterases—one of the termination enzymes described herein—and the reverse beta oxidation pathway has also been achieved in yeast. Other species include but are not limited to *Candida*, *Aspergillus*, *Arxula adenivorans*, *Candida boidinii*, *Hansenula polymorpha* (*Pichia angusta*), *Kluyveromyces lactis*, *Pichia pastoris*, and *Yarrowia lipolytica*, to name a few.

[0035] It is also possible to genetically modify many species of algae, including e.g., *Spirulina*, *Apergillus*, *Chlamydomonas*, *Laminaria japonica*, *Undaria pinnatifida*, *Porphyra*, *Eucheuma*, *Kappaphycus*, *Gracilaria*, *Monostroma*, *Enteromorpha*, *Arthrospira*, *Chlorella*, *Dunaliella*, *Aphanizomenon*, *Isochrysis*, *Pavlova*, *Phaeodactylum*, *Ulkenia*, *Haematococcus*, *Chaetoceros*, *Nannochloropsis*, *Skeletonema*, *Thalassiosira*, and *Laminaria japonica*, and the like. Indeed, the microalga *Pavlova lutheri* is already being used as a source of economically valuable docosahexaenoic (DHA) and eicosapentaenoic acids (EPA), and *Cryptothecodinium cohnii* is the heterotrophic algal species that is currently used to produce the DHA used in many infant formulas.

[0036] Furthermore, a number of databases include vector information and/or a repository of vectors and can be used to choose vectors suitable for the chosen host species. See e.g., AddGene.org which provides both a repository and a searchable database allowing vectors to be easily located and obtained from colleagues. See also Plasmid Information Database (PlasmID) and DNASU having over 191,000 plasmids. A collection of cloning vectors of *E. coli* is also kept at the National Institute of Genetics as a resource for the biological research community. Furthermore, vectors (including particular ORFs therein) are usually available from colleagues.

[0037] The enzymes can be added to the genome or via expression vectors, as desired. Preferably, multiple enzymes are expressed in one vector or multiple enzymes can be combined into one operon by adding the needed signals between coding regions. Further improvements can be had by overexpressing one or more, or even all of the enzymes, e.g., by adding extra copies to the cell via plasmid or other vector. Initial experiments may employ expression plasmids hosting 3 or more ORFs for convenience, but it may be preferred to insert operons or individual genes into the genome for long term stability.

[0038] Still further improvements in yield can be had by reducing competing pathways, such as those pathways for making e.g., acetate, formate, ethanol, and lactate, and it is already well known in the art how to reduce or knockout these pathways. See e.g., the Rice University's patent portfolio by Ka-Yiu San and George Bennett (U.S. Pat. Nos. 7,569,380, 7,262,046, 8,962,272, 8,795,991) and patents by these inventors (U.S. Pat. Nos. 8,129,157 and 8,691,552) (each incorporated by reference herein in its entirety for all purposes). Many others have worked in this area as well.

[0039] In calculating “% identity” the unaligned terminal portions of the query sequence are not included in the calculation. The identity is calculated over the entire length of the reference sequence, thus short local alignments with a query sequence are not relevant (e.g., % identity=number of aligned residues in the query sequence/length of reference sequence). Alignments are performed using BLAST homology alignment as described by Tatusova T A & Madden T L (1999) *FEMS Microbiol. Lett.* 174:247-250, and available through the NCBI website. The default parameters were used, except the filters were turned OFF.

[0040] “Operably associated” or “operably linked”, as used herein, refer to functionally coupled nucleic acid or amino acid sequences.

[0041] “Recombinant” is relating to, derived from, or containing genetically engineered material. In other words, the genetics of an organism was intentionally manipulated by the hand of man in some way.

[0042] “Reduced activity” is defined herein to be at least a 75% reduction in protein activity, as compared with an appropriate control species (e.g., the wild type gene in the same host species). Preferably, at least 80, 85, 90, 95% reduction in activity is attained, and in the most preferred embodiment, the activity is eliminated (100%). Proteins can be inactivated with inhibitors, by mutation, or by suppression of expression or translation, by knock-out, by adding stop codons, by frame shift mutation, and the like. All reduced activity genes or proteins are signified herein by “-”.

[0043] By “null” or “knockout” what is meant is that the mutation produces undetectable active protein. A gene can

be completely (100%) reduced by knockout or removal of part of all of the gene sequence. Use of a frame shift mutation, early stop codon, point mutations of critical residues, or deletions or insertions, and the like, can also completely inactivate (100%) gene product by completely preventing transcription and/or translation of active protein. All null mutants herein are signified by Δ .

[0044] “Overexpression” or “overexpressed” is defined herein to be at least 150% of protein activity as compared with an appropriate control species, or any detectable expression in a species that lacks the activity altogether. Preferably, the activity is increased 100-500% or even ten fold. Overexpression can be achieved by mutating the protein to produce a more active form or a form that is resistant to inhibition, by removing inhibitors, or adding activators, and the like. Overexpression can also be achieved by removing repressors, adding multiple copies of the gene to the cell, or up-regulating the endogenous gene, and the like. All overexpressed genes or proteins are signified herein by “+”.

[0045] In certain species it is possible to genetically engineer the endogenous protein to be overexpressed by changing the regulatory sequences or removing repressors. However, overexpressing the gene by inclusion on selectable plasmids or other vectors that exist in hundreds of copies in the cell may be preferred due to its simplicity and ease of exerting external controls, although permanent modifications to the genome may be preferred in the long term for stability reasons.

[0046] The term “endogenous” or “native” means that a gene originated from the species in question, without regard to subspecies or strain, although that gene may be naturally or intentionally mutated, or placed under the control of a promoter that results in overexpression or controlled expression of said gene. Thus, genes from *Clostridia* would not be endogenous to *Escherichia*, but a plasmid expressing a gene from *E. coli* or would be considered to be endogenous to any genus of *Escherichia*, even though it may now be overexpressed.

[0047] “Expression vectors” are used in accordance with the art accepted definition of a plasmid, virus or other propagatable sequence designed for protein expression in cells. There are thousands of such vectors commercially available, and typically each has an origin of replication (ori); a multiple cloning site; a selectable marker; ribosome binding sites; a promoter and often enhancers; and the needed termination sequences. Most expression vectors are inducible, although constitutive expressions vectors also exist.

[0048] As used herein, “inducible” means that gene expression can be controlled by the hand of man, by adding e.g., a ligand to induce expression from an inducible promoter. Exemplary inducible promoters include the lac operon, inducible by IPTG, the yeast AOX1 promoter inducible with methanol, the strong LAC4 promoter inducible with lactate, and the like. Low level of constitutive protein synthesis may occur even in expression vectors with tightly controlled promoters.

[0049] As used herein, an “integrated sequence” means the sequence has been integrated into the host genome, as opposed to being maintained on an expression vector. It will still be expressible, and preferably is inducible as well.

[0050] The use of the word “a” or “an” when used in conjunction with the term “comprising” in the claims or the specification means one or more than one, unless the context dictates otherwise.

[0051] The term “about” means the stated value plus or minus the margin of error of measurement or plus or minus 10% if no method of measurement is indicated.

[0052] The use of the term “or” in the claims is used to mean “and/or” unless explicitly indicated to refer to alternatives only or if the alternatives are mutually exclusive.

[0053] The terms “comprise”, “have”, “include” and “contain” (and their variants) are open-ended linking verbs and allow the addition of other elements when used in a claim.

[0054] The phrase “consisting of” is closed, and excludes all additional elements.

[0055] The phrase “consisting essentially of” excludes additional material elements, but allows the inclusions of non-material elements that do not substantially change the nature of the invention, such as instructions for use, buffers, background mutations that do not effect the invention, and the like.

[0056] The following abbreviations are used herein:

ABBREVIATION	TERM
ACP	Acyl carrier protein
ATP	Adenosine triphosphate
Box-R	Beta oxidation pathway in reverse
CoA	Coenzyme A
FAS	Fatty acid biosynthesis
HACD	Hydroxyacyl-CoA dehydrogenases
ECH	Enoyl-CoA hydratase
ECR	Enoyl-CoA reductase
HACL	2-hydroxyacyl-CoA lyase

BRIEF DESCRIPTION OF THE DRAWINGS

[0057] FIG. 1A and FIG. 1B: Platform for the synthesis of alpha-functionalized carboxylic acids, alcohols and amines. Acyl-CoA primer, which is either unsubstituted or functionalized, and alpha-functionalized extender unit are mainly activated from their acid form, which can be either supplemented in the media or derived from carbon sources. Primer and extender unit can also be derived from carbon sources without the need to generate their acid forms. The platform is composed of thiolases, dehydrogenases, dehydratases and reductases. Thiolases catalyze a condensation between acyl-CoA primer and alpha-functionalized acyl-CoA extender and generates alpha-functionalized β -keto acyl-CoA. Dehydrogenases convert alpha-functionalized β -keto acyl-CoA to alpha-functionalized β -hydroxy acyl-CoA. FIG. 1B shows dehydratases convert alpha-functionalized β -hydroxy acyl-CoA to alpha-functionalized enoyl-CoA. Reductases convert alpha-functionalized enoyl-CoA to alpha-functionalized acyl-CoA. Iterative operation can be realized by using alpha-functionalized acyl-CoA as primer and either acetyl-CoA or alpha-functionalized acetyl-CoA as extender unit in subsequent turns of the platform. Termination pathways starting from four alpha-functionalized CoA thioester intermediates terminate the platform and generate various alpha-functionalized carboxylic acids, alcohols and amines with different β -reduction degrees. There are three types of termination pathways: thioesterase/CoA-transferase/phosphotransacylase+kinase, which generates carboxylic acids; acyl-CoA reductase and alcohol dehydrogenase which gen-

erate alcohols; acyl-CoA reductase and transaminase which generate amine. R_1 and R_2 mean functionalized group from primer and extender unit respectively. Dashed line means multiple reaction steps or iteration.

[0058] FIG. 2A and FIG. 2B: Proposed platform depicted in FIGS. 1A and 1B and its products utilizing propionyl-CoA as the extender unit (R_2 in FIGS. 1A and 1B=—CH₃).

[0059] FIG. 3: Example pathway of synthesis of tiglic acid (trans-2-methyl-2-butenic acid) and 2-methylbutyric acid through the proposed platform with acetyl-CoA as the primer and propionyl-CoA as the extender unit. Propionyl-CoA is activated by Pct from propionic acid (Step 1). The platform is composed of thiolase FadAx, which catalyzes the condensation between primer acetyl-CoA and extender unit propionyl-CoA to 2-methyl acetoacetyl-CoA (Step 2); dehydrogenase FadB2x, which converts 2-methyl acetoacetyl-CoA to 2-methyl-3-hydroxybutyryl-CoA (Step 3); dehydratase FadB1x, which converts 2-methyl-3-hydroxybutyryl-CoA to tiglyl-CoA (Step 4); reductase FabI, which reduces tiglyl-CoA to 2-methylbutyryl-CoA (Step 5). Termination reactions by endogenous thioesterases from tiglyl-CoA (Step 6) and 2-methylbutyryl-CoA (Step 7) finally generate products tiglic acid and 2-methylbutyric acid.

[0060] FIG. 4: Example pathway of synthesis of trans-2-methyl-2-pentenoic acid and 2-methylvaleric acid through the proposed platform with propionyl-CoA as the primer and the extender unit. Propionyl-CoA is activated by Pct from propionic acid (Step 1). The platform is composed of thiolase FadAx, which catalyzes the condensation between two molecules of propionyl-CoA to 2-methyl-3-oxopentanoyl-CoA (Step 2); dehydrogenase FadB2x, which converts 2-methyl-3-oxopentanoyl-CoA to 2-methyl-3-hydroxypentanoyl-CoA (Step 3); dehydratase FadB1x, which converts 2-methyl-3-hydroxypentanoyl-CoA to 2-methyl-2-pentenoyl-CoA (Step 4); reductase FabI, which reduces 2-methyl-2-pentenoyl-CoA to 2-methylvaleryl-CoA (Step 5). Termination reactions by endogenous thioesterases from 2-methyl-2-pentenoyl-CoA (Step 6) and 2-methylvaleryl-CoA (Step 7) finally generate products 2-methyl-2-pentenoic acid and 2-methylvaleric acid.

[0061] FIG. 5: Titers of alpha-methylated products synthesized through the utilization of propionyl-CoA as the extender unit with either acetyl-CoA or propionyl-CoA priming. These products were produced from the *E. coli* strain overexpressing enzymes catalyzing Steps 1-5 depicted in FIG. 3-4. JC01(DE3), an *E. coli* strain deficient of mixed-acid fermentations, served as the host strain. The engineered strains were grown for 48 hours under 37° C. in 20 mL LB-like MOPS media supplemented with 20 g/L glycerol and 20 mM propionic acid.

[0062] FIG. 6: Pathway for the improved production of tiglic acid through the proposed platform with acetyl-CoA as the primer and propionyl-CoA as the extender unit. Propionyl-CoA is activated by Pct from propionic acid (Step 1). Thiolase FadAx condenses acetyl-CoA and propionyl-CoA to 2-methyl acetoacetyl-CoA (Step 2). Dehydrogenase FadB2x converts 2-methyl acetoacetyl-CoA to 2-methyl-3-hydroxybutyryl-CoA (Step 3). Dehydratase FadB1x converts 2-methyl-3-hydroxybutyryl-CoA to tiglyl-CoA (Step 4). Finally, thioesterase YdiI can remove the CoA from tiglyl-CoA to generate the product tiglic acid (Step 5).

[0063] FIG. 7: Results of improvement of tiglic acid production by removal of overexpression of FabI (ECR),

addition of overexpression of YdiI (a thioesterase) and usage of JST06(DE3) as the host strain. JST06(DE3) is an *E. coli* strain deficient of mixed-acid fermentations, thioesterases. The engineered strains were grown for 48 h at 37° C. in 20 mL LB-like MOPS media supplemented with 20 g/L glycerol and 20 mM propionic acid.

[0064] FIG. 8: Time course for tiglic acid production from JST06(DE3) strain overexpressing Pct, FadAx, FadB2x, FadB1x and YdiI in a fermentation conducted in a controlled bioreactor. The fermentation was performed under 37° C. in LB-like MOPS media supplemented with 30 g/L glycerol, and 20 mM propionic acid which was added at 0, 24, and 48 h.

[0065] FIG. 9A and FIG. 9B: Proposed platform depicted in FIGS. 1A and 1B and its products utilizing glycolyl-CoA as the extender unit (R_2 in FIGS. 1A and 1B=—OH).

[0066] FIG. 10: Example pathway of synthesis of 2,3-dihydroxy-butyric acid through the proposed platform with acetyl-CoA as the primer and propionyl-CoA as the extender unit. Glycolyl-CoA is activated by Pct from glycolic acid (Step 1). Then, condensation by thiolase BktB converts glycolyl-CoA and acetyl-CoA to 2-hydroxy acetoacetyl-CoA (Step 2). Dehydrogenase PhaB converts 2-hydroxy acetoacetyl-CoA to 2,3-dihydroxy-butyryl-CoA (Step 3). CoA removal by endogenous thioesterases convert 2,3-dihydroxy-butyryl-CoA to the product 2,3-dihydroxy-butyric acid (Step 4).

[0067] FIG. 11: Peak of product 2,3-dihydroxy-butyric acid in the GC-MS chromatogram of the fermentation sample from MG1655(DE3) Δ glcD (pET-P1-bktB-phaB-P2-phaJ) (pCDF-P1-pct-P2-tdTER). The strain was grown in 50 mL LB media supplemented with 10 g/L glucose and 40 mM glycolate for 96 hours under 30° C. in 250 mL flask.

[0068] FIG. 12: Derivatization pathway of product 2-hydroxy acid and intermediate 2-hydroxyacyl-CoA of the proposed platform utilizing glycolyl-CoA as the extender unit depicted in FIG. 3, to a primary alcohol product. 2-hydroxyacyl-CoA can be degraded to primary aldehyde and formyl-CoA by 2-hydroxyacyl-CoA lyase. 2-hydroxy acid can be converted to α -keto acid by keto-dehydrogenase and α -keto acid can be decarboxylated to primary aldehyde by α -keto acid to primary aldehyde. Primary aldehyde is finally reduced to primary alcohol by alcohol dehydrogenase.

[0069] FIG. 13: Vector map of pCDFDuet-1-P1-ntH6-HACL1 for overexpression and purification of codon-optimized 2-hydroxyacyl-CoA lyase HACL1 from *Homo sapiens* in *E. coli*.

[0070] FIG. 14: SDS-PAGE analysis result of overexpression of *Homo sapiens* HACL1 in *E. coli*.

[0071] FIG. 15: Vector map of pYES260-HACL1-SCopt for overexpression and purification of codon-optimized 2-hydroxyacyl-CoA lyase HACL1 from *Homo sapiens* in *Saccharomyces cerevisiae*.

[0072] FIG. 16: SDS-PAGE analysis result of overexpression and purification of *Homo sapiens* HACL1 in *S. cerevisiae*.

[0073] FIG. 17: GC-FID chromatograms of pentadecanal content in HACL1 degradative reaction (forward reaction) mixtures after extraction with hexane. HACL1 was expressed and purified from *S. cerevisiae*. Top: pentadecanal standard; Middle: HACL1 assay sampled; Bottom: no enzyme control. In samples containing HACL1, a pentadecanal peak is seen, while there is no peak in the sample in which enzyme was omitted.

[0074] FIG. 18: GC-FID chromatograms of pentadecanal content demonstrating HACL1 activity in *E. coli* BL21 (DE3) crude extract. The peak of pentadecanal is shown in the square.

[0075] FIG. 19A and FIG. 19B: Proposed platform depicted in FIGS. 1A and 1B and its products utilizing phenylacetyl-CoA as the extender unit (R_2 in FIGS. 1A and 1B=—Ph).

[0076] FIG. 20A and FIG. 20B: Proposed platform depicted in FIG. 1 and its products utilizing phenylacetyl-CoA as the extender unit (R_2 in FIGS. 1A and 1B=—NH₂).

[0077] FIG. 21. A partial listing of embodiments of the invention, any one or more of which can be combined with any other.

DETAILED DESCRIPTION

[0078] This disclosure generally relates to the use of microorganisms to make alpha-functionalized chemicals and fuels, (e.g. alpha-functionalized carboxylic acids, alcohols, hydrocarbons, amines, and their beta-, and omega-functionalized derivatives), by utilizing a novel iterative carbon chain elongation pathway that uses functionalized extender units to grow a carbon chain by two carbon units.

[0079] The core enzymes in the pathway include thiolase, dehydrogenase, dehydratase and reductase. Native or engineered thiolases catalyze the condensation of either unsubstituted or functionalized acyl-CoA primers with an alpha-functionalized acetyl-CoA as the extender unit to generate alpha-functionalized β -keto acyl-CoA. Dehydrogenase converts alpha-functionalized β -keto acyl-CoA to alpha-functionalized β -hydroxy acyl-CoA. Dehydratase converts alpha-functionalized β -hydroxy acyl-CoA to alpha-functionalized enoyl-CoA. Reductase converts alpha-functionalized enoyl-CoA to alpha-functionalized acyl-CoA.

[0080] The platform can be operated in an iterative manner (i.e. multiple turns) by using the resulting alpha-functionalized acyl-CoA as primer and the aforementioned omega-functionalized extender unit in subsequent turns of the cycle. Various termination pathways (FIGS. 1A and 1B and Table 4) acting on any of the four alpha-functionalized CoA thioester intermediates terminate the platform and generate various alpha-functionalized carboxylic acids, alcohols and amines with different β -reduction degrees.

[0081] Thioesterase or CoA transferase or phosphotransacylase+carboxylate kinase can terminate the platform by converting the alpha-functionalized acyl-CoAs to alpha-functionalized carboxylic acids. If alpha-functionalized carboxylic acids has keto group at the beta-site, it can then be converted to ketone through reactions by β -keto acid decarboxylase. Acyl-CoA reductases can terminate the platform by converting the alpha-functionalized acyl-CoAs to alpha-functionalized aldehydes. Alpha-functionalized aldehydes can then be converted to alpha-functionalized alcohols and alpha-functionalized amines through reactions by alcohol dehydrogenase and transaminase respectively.

[0082] This disclosure also relates to a novel primary alcohol synthesis incorporating the proposed iterative platform using glycolyl-CoA (alpha-hydroxy acetyl-CoA) as the extender unit. When the platform uses glycolyl-CoA as the extender unit, it generates alpha-hydroxyacyl-CoA, which can be converted to primary alcohol by termination pathways selected from: a) 2-hydroxyacyl-CoA lyase (HACL) that converts alpha-hydroxyacyl-CoA to primary aldehyde with one less carbon and formyl-CoA, and alcohol dehydrogenase subsequently converts the primary aldehyde to primary alcohol; b) acid-forming termination enzyme selected from thioesterase, CoA transferase and phosphotransacylase+carboxylate kinase that converts alpha-hydroxyacyl-CoA to alpha-hydroxy acid, keto-dehydrogenase that converts alpha-hydroxy acid to alpha-keto acid, alpha-keto acid decarboxylase that converts alpha-keto acid to primary aldehyde with one less carbon and alcohol dehydrogenase subsequently converts the primary aldehyde to primary alcohol.

[0083] Many examples of thiolase enzymes which can potentially catalyze the non-decarboxylative condensation of an acyl-CoA primer and acetyl-CoA extender unit are provided herein and Table 1 provides several additional examples which can also serve as templates for engineered variants:

TABLE 1

Example Thiolase Enzymes (EC Number 2.3.1.-)	
Source organism and gene name	Protein Accession Numbers
<i>E. coli</i> atoB	NP_416728.1
<i>E. coli</i> yqeF	NP_417321.2
<i>E. coli</i> fadA	YP_026272.1
<i>E. coli</i> fadI	NP_416844.1
<i>Streptomyces collinus</i> fadA	Q93C88
<i>Ralstonia eutropha</i> bktB	AAC38322.1
<i>Pseudomonas</i> sp. Strain B13 catF	AAL02407.1
<i>E. coli</i> paaJ	NP_415915.1
<i>Pseudomonas putida</i> pcaF	AAA85138.1
<i>Rhodococcus opacus</i> pcaF	YP_002778248.1
<i>Streptomyces</i> sp. pcaF	AAD22035.1
<i>Ralstonia eutropha</i> phaA	AEI80291.1
<i>Clostridium acetobutylicum</i> thIA	AAC26023.1
<i>Clostridium acetobutylicum</i> thIB	AAC26026.1

[0084] This technology takes the above thiolase initiated pathway one step further to make alpha functionalized products. The method entails developing a new pathway that is based on native or engineered thiolases capable of catalyzing the condensation of either unsubstituted or functionalized acyl-CoA primers with an omega-functionalized acetyl-CoA as the extender unit. This has been reported in neither the scientific, peer-reviewed literature nor the patent literature.

[0085] Materials that can be used with the invention include those in Tables 2-5 below.

TABLE 2

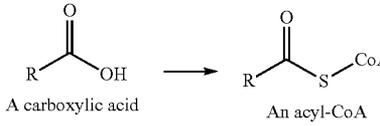
Activation enzymes							
Reaction	Illustration	EC Numbers	Enzyme names	Source organism and gene name	Protein Accession Numbers		
Carboxylic acid → Acyl-CoA (including acyl-CoA primer, and α-functionalized acetyl-CoA acting as the extender unit)		6.2.1.-	Acyl-CoA synthetase	<i>E. coli paaK</i>	NP_415916.1		
				<i>E. coli sucCD</i>	NP_415256.1		
				<i>E. coli fadK</i>	NP_415257.1		
				<i>E. coli fadD</i>	NP_416216.4		
				<i>E. coli prpE</i>	NP_416319.1		
				<i>E. coli menE</i>	NP_414869.1		
				<i>Penicillium chrysogenum phl</i>	NP_416763.1		
				<i>Salmonella typhimurium LT2 prpE</i>	CAJ15517.1		
				<i>Bacillus subtilis bioW</i>	AAL19325.1		
				<i>Cupriavidus basilensis hmjD</i>	AAC00261.1		
				<i>Rhodopseudomonas palustris badA</i>	ADE20402.1		
				<i>R. palustris hbaA</i>	CAJ18317.1		
				<i>Pseudomonas aeruginosa PAO1 pqsA</i>	CAE26113.1		
				<i>Arabidopsis thaliana 4cl</i>	NP_249687.1		
				2.8.3-	CoA transferase	<i>E. coli atoD</i>	Q42524.1
						<i>E. coli atoA</i>	NP_416725.1
						<i>E. coli scpC</i>	NP_416726.1
						<i>Clostridium kluyveri cat1</i>	NP_417395.1
						<i>Clostridium kluyveri cat2</i>	AAA92346.1
						<i>Clostridium acetobutylicum cfAB</i>	AAA92344.1
<i>Pseudomonas putida pcalJ</i>	NP_149326.1						
<i>Megasphaera elsdenii pct</i>	NP_149327.1						
<i>Acidaminococcus fermentans gctAB</i>	NP_746081.1						
<i>Acetobacter acetii aarC</i>	NP_746082.1						
<i>E. coli ydiF</i>	WP_014015705.1						
2.3.1 -;	Phospho-trans-acylase +	<i>Clostridium acetobutylicum ptb</i>	NP_349676.1				
		<i>Enterococcus faecalis ptb</i>	AAD55374.1				
2.7.2.15	Carboxylate kinase	<i>Salmonella enterica pduL</i>	AAD39011.1				
		<i>Clostridium acetobutylicum buk</i>	AAK81015.1				
		<i>Enterococcus faecalis buk</i>	AAD55375.1				
		<i>Salmonella enterica pduW</i>	AAD39021.1				

TABLE 3

Reactions of the platform						
Reaction	Illustration	EC Numbers	Enzyme names	Source organism and gene name	Protein Accession Numbers	
Acyl-CoA + α -functionalized acetyl-CoA \rightarrow α -functionalized β -ketoacyl-CoA		2.3.1.-	Thiolase	<i>E. coli atoB</i> <i>E. coli yqeF</i> <i>E. coli fadA</i> <i>E. coli fadI</i> <i>Ralstonia eutropha bktB</i> <i>Pseudomonas</i> sp. Strain B13 <i>catF</i> <i>E coli paaJ</i> <i>Pseudomonas putida pcaF</i> <i>Rhodococcus opacus pcaF</i> <i>Streptomyces</i> sp. <i>pcaF</i> <i>Ralstonia eutropha phaA</i> <i>Clostridium acetobutylicum thlA</i> <i>Clostridium acetobutylicum thlB</i> <i>Pseudomonas putida fadA</i> <i>P. putida fadAx</i> <i>Acinetobacter</i> sp. ADP1 <i>dcaF</i> <i>E. coli paaJ</i>	NP_416728.1 NP_417321.2 YP_026272.1 NP_416844.1 AAC38322.1 AAL02407.1 NP_415915.1 AAA85138.1 YP_002778248.1 AAD22035.1 AEI80291.1 AAC26023.1 AAC26026.1 AAK18168.1 AAK18171.1 CAG68532.1 NP_415915.1	
α -functionalized β -ketoacyl-CoA \rightarrow α -functionalized β -hydroxyacyl-CoA		1.1.1.35; 1.1.1.36	Hydroxyacyl-CoA dehydrogenase	<i>E. coli fadB</i> <i>E. coli fadJ</i> <i>E. coli paaH</i> <i>P. putida fadB</i> <i>P. putida fadB2x</i> <i>Acinetobacter</i> sp. ADP1 <i>dcaF</i> <i>Ralstonia eutrophus phaB</i> <i>Clostridium acetobutylicum hbd</i> <i>E. coli fabG</i>	NP_418288.1 NP_416843.1 NP_415913.1 AAK18167.2 AAK18170.1 CAG68533.1 P14697.1 AAA95971.1 NP_415611.1	
α -functionalized β -hydroxyacyl-CoA \rightarrow α -functionalized enoyl-CoA		4.2.1.17; 4.2.1.119	enoyl-CoA hydratase	<i>E. coli fadB</i> <i>E. coli fadJ</i> <i>E. coli paaF</i> <i>P. putida fadB</i> <i>P. putida fadB1x</i> <i>Acinetobacter</i> sp. ADP1 <i>dcaE</i> <i>Clostridium acetobutylicum crt</i> <i>Aeromonas caviae phaJ</i> <i>E. coli fabA</i> <i>E. coli fabZ</i>	NP_418288.1 NP_416843.1 NP_415911.1 AAK18167.2 AAK18173.1 CAG68535.1 AAA95967.1 O32472.1 NP_415474.1 NP_414722.1	
α -functionalized enoyl-CoA \rightarrow α -functionalized acyl-CoA		1.3.1.44	enoyl-CoA reductase	<i>Euglena gracilis TER</i> <i>Treponema denticola TER</i> <i>Clostridium acetobutylicum TER</i>	Q5EU90.1 4GGO_A 4EUH_A	

TABLE 3-continued

Reactions of the platform						
Reaction	Illustration	EC Numbers	Enzyme names	Source organism and gene name	Protein Accession Numbers	
			enoyl-[acyl-carrier-protein] reductase	<i>E. coli fabI</i> <i>Enterococcus faecalis fabK</i> <i>Bacillus subtilis fabL</i> <i>Vibrio cholerae fabV</i>	NP_415804.1 NP_816503.1 KFK80655.1 ABX38717.1	
			acyl-CoA dehydrogenase	<i>E. coli fadE</i> <i>E. coli ydiO</i>	NP_414756.2 NP_416210.4	

② indicates text missing or illegible when filed

TABLE 4

Termination Pathways						
Reaction	Illustration	EC Numbers	Enzyme names	Source organism and gene name	Protein Accession Numbers	
Acyl-CoA → Carboxylic acid	<p>An acyl-CoA → A carboxylic acid</p>	3.1.2 -	Thioesterase	<i>E. coli tesA</i> <i>E. coli tesB</i> <i>E. coli yciA</i> <i>E. coli fadM</i> <i>E. coli ydiI</i> <i>E. coli ybgC</i> <i>E. coli paal</i> <i>Mus musculus acot8</i> <i>Lycopersicon hirsutum f glabratum mks2</i> <i>Alcanivorax borkumensis tesB2</i> <i>Fibrobacter succinogenes Fs2108</i> <i>Prevotella ruminicola Pr655</i> <i>Prevotella ruminicola Pr1687</i>	NP_415027.1 NP_414986.1 NP_415769.1 NP_414977.1 NP_416201.1 NP_415264.1 NP_415914.1 P58137.1 ADK38536.1 YP_692749.1 YP_005822012.1 YP_003574018.11 YP_003574982.1	
		2.8.3-	CoA transferase	<i>E. coli atoD</i> <i>E. coli atoA</i> <i>E. coli scpC</i> <i>Clostridium kluyvericat1</i> <i>Clostridium kluyvericat2</i> <i>Clostridium acetobutylicum ctfAB</i> <i>Pseudomonas putida pcalJ</i> <i>Megasphaera elsdeni pct</i> <i>Acidaminococcus fermentans gctAB</i> <i>Acetobacter acetii aarC</i> <i>E. coli ydiF</i>	NP_416725.1 NP_416726.1 NP_417395.1 AAA92346.1 AAA92344.1 NP_149326.1, NP_149327.1 NP_746081.1 NP_746082.1 WP_014015705.1 CAA57199.1 CAA57200.1 AGG68319.1 NP_416209.1	
		2.3.1 -; 2.7.2.1; 2.7.2.15	Phosphotrans-acylase + Carboxylate kinase	<i>Clostridium acetobutylicum ptb</i> <i>Enterococcus faecalis ptb</i> <i>Salmonella enterica pduL</i> <i>Clostridium acetobutylicum buk</i>	NP_349676.1 AAD55374.1 AAD39011.1 AAK81015.1	

TABLE 4-continued

Termination Pathways						
Reaction	Illustration	EC Num- bers	Enzyme names	Source organism and gene name	Protein Accession Numbers	
Acyl-CoA → Aldehyde	<p>An acyl-CoA → An aldehyde</p>	1.2.1.10	Aldehyde forming CoA reductase	<i>Enterococcus faecalis buk</i> <i>Salmonella enterica pduW</i> <i>Acinetobacter calcoaceticus acr1</i> <i>Acinetobacter sp Strain M-1 acrM</i> <i>Clostridium beijerinckii ald</i> <i>E. coli eutE</i> <i>Salmonella enterica eutE</i> <i>Marinobacter aquaeolei VT8 maqu_2507</i> <i>E. coli mhpF</i> <i>Clostridium kluyveri sucD</i>	AAD55375.1 AAD39021.1 AAC45217.1 BAB85476.1 AAT66436.1 NP_416950.1 AAA80209.1 YP_959769.1 NP_414885.1 EDK35023.1	
Aldehyde → Alcohol	<p>An aldehyde → An alcohol</p>	1.1.1.-	Alcohol dehydrogenase	<i>E. coli beta</i> <i>E. coli dkgA</i> <i>E. coli eutG</i> <i>E. coli fucO</i> <i>E. coli ucpA</i> <i>E. coli yahK</i> <i>E. coli ybbO</i> <i>E. coli ybdH</i> <i>E. coli yiaY</i> <i>E. coli yjgB</i> <i>Marinobacter aquaeolei VT8 maqu_2507</i> <i>Saccharomyces cerevisiae ADH6</i> <i>Clostridium kluyveri 4hbD</i> <i>Acinetobacter sp. SE19 chnD</i>	NP_414845.1 NP_417485.4 NP_416948.4 NP_417279.2 NP_416921.4 NP_414859.1 NP_415026.1 NP_415132.1 YP_026233.1 NP_418690.4 YP_959769.1 Q04894.1 EDK35022.1 AAG10028.1	
Aldehyde → Amine	<p>An aldehyde → An amine</p>	2.6.1.-	Transaminase	<i>Arabidopsis thaliana At3g22200</i> <i>Alcaligenes denitrificans AptA</i> <i>Bordetella bronchiseptica BB0869</i> <i>Bordetella parapertussis BPP0784</i> <i>Brucella melitensis BAWG_0478</i> <i>Burkholderia pseudomallei BP1026B_10669</i> <i>Chromobacterium violaceum CP2025</i> <i>Oceanicola granulosus OG2516_07293</i> <i>Paracoccus denitrificans PD1222</i> <i>Pden_3984</i> <i>Pseudogulbenkiania ferrooxidans ω-TA</i> <i>Pseudomonas putida ω-TA</i> <i>Ralstonia solanacearum ω-TA</i>	NP_001189947.1 AAP92672.1 WP_015041039.1 WP_0109276831 EEW88370.1 AFI65333.1 AAQ59697.1 WP_007254984.1 ABL72050.1 WP_008952788.1 P28269.1 YP_002258353.1	

TABLE 4-continued

Termination Pathways					
Reaction	Illustration	EC Num- bers	Enzyme names	Source organism and gene name	Protein Accession Numbers
				<i>Rhizobium meliloti</i> SMc01534	NP_386510.1
				<i>Vibrio fluvialis</i> ω-TA	AEA39183.1
				<i>Mus musculus</i> <i>abaT</i>	AAH58521.1
				<i>Flavobacterium lutescens</i> <i>lat</i>	BAB13756.1
				<i>Streptomyces clavuligerus</i> <i>lat</i>	AAB39899.1
				<i>E. coli</i> <i>gabT</i>	YP_490877.1
				<i>E. coli</i> <i>puuE</i>	NP_415818.1
				<i>E. coli</i> <i>ygjG</i>	NP_417544.5
β-keto acid → ketone	<p>A α-functionalized β-keto acid</p> <p>A ketone</p>	4.1.1.56;	β-keto acid decarboxylase	<i>Lycopersicon hirsutum</i> <i>f glabratum</i> <i>mks1</i> <i>Clostridium acetobutylicum</i> <i>adc</i>	ADK38535.1 AAA63761.1

TABLE 5

Enzymes for derivatization of 2-hydroxy acid to primary alcohol					
Reaction	Illustration	EC Num- bers	Enzyme names	Source organism and gene name	Protein Accession Numbers
2-hydroxy acid → α-keto acid	<p>A 2-hydroxy acid</p> <p>An α-keto acid</p>	1.1.1-	Keto-dehydrogenase	<i>Clostridium beijerinckii</i> <i>adh</i> <i>E. coli</i> <i>serA</i> <i>Gordonia</i> <i>sp. TY-5</i> <i>adh1</i> <i>Gordonia</i> <i>sp. TY-5</i> <i>adh2</i> <i>Gordonia</i> <i>sp. TY-5</i> <i>adh3</i> <i>Rhodococcus ruber</i> <i>adh-A</i> <i>Acidaminococcus fermentans</i> <i>hgdH</i> <i>E. coli</i> <i>IdhA</i> <i>E. coli</i> <i>IldD</i> <i>E. coli</i> <i>leuB</i>	AAA23199.2 NP_417388.1 BAD03962.1 BAD03964.1 BAD03961.1 WP_043801412.1 ADB47349.1 NP_415898.1 NP_418062.1 NP_414615.4
α-keto acid → primary aldehyde	<p>An α-keto acid</p> <p>A primary aldehyde</p>	4.1.1.1	α-keto acid decarboxylase	<i>Lactococcus lactis</i> <i>kivd</i> <i>Saccharomyces cerevisiae</i> <i>PDC1</i> <i>S. cerevisiae</i> <i>PDC5</i> <i>S. cerevisiae</i> <i>PDC6</i> <i>S. cerevisiae</i> <i>ARO10</i> <i>S. cerevisiae</i> <i>THI3</i> <i>Zymomonas mobilis</i> <i>pdc</i>	AIS03677.1 CAA97573.1 CAA97705.1 CAA97089.1 NP_010668.3 CAA98646.1 ADK13058.1

TABLE 5-continued

Enzymes for derivatization of 2-hydroxy acid to primary alcohol						
Reaction	Illustration	EC Numbers	Enzyme names	Source organism and gene name	Protein Accession Numbers	
Primary aldehyde → Primary alcohol	<p>A primary aldehyde</p>	1.1.1.-	Alcohol dehydrogenase	<i>E. coli betA</i> <i>E. coli dkgA</i> <i>E. coli eutG</i> <i>E. coli fucO</i> <i>E. coli ucpA</i> <i>E. coli yahK</i> <i>E. coli ybbO</i> <i>E. coli ybdH</i> <i>E. coli yiaY</i> <i>E. coli yjgB</i> <i>Saccharomyces cerevisiae ADH6</i> <i>Clostridium kluyveri 4hbD</i> <i>Acinetobacter</i> sp. SE19 <i>chmD</i>	NP_414845.1 NP_417485.4 NP_416948.4 NP_417279.2 NP_416921.4 NP_414859.1 NP_415026.1 NP_415132.1 YP_026233.1 NP_418690.4 Q04894.1 EDK35022.1 AAG10028.1	
2-hydroxyacyl-CoA → primary aldehyde + formyl-CoA	<p>A 2-hydroxyacyl-CoA</p> <p>A formyl-CoA</p> <p>A primary aldehyde</p>	4.1.-.	2-hydroxyacyl-CoA lyase	<i>Homo sapiens hac1</i> <i>Rattus norvegicus hac1</i> <i>Dictyostelium discoideum hac1</i> <i>Mus musculus hac1</i>	Q9UJ83 Q8CHM7 Q54DA9 Q9QXE0	

[0086] All strains used in this study are listed in Table 6. Gene deletions were performed using P1 phage transduction with single-gene knockout mutants from the National BioResource Project (NIG, Japan) as the specific deletion donor. The λ DE3 prophage, carrying the T7 RNA polymerase gene and *lacIq*, was integrated into the chromosome through λ DE3 lysogenization kit (Novagen, Darmstadt, Germany). All strains were stored in 32.5% glycerol stocks at -80° C. Plates were prepared using LB medium containing 1.5% agar, and appropriate antibiotics were included at the following concentrations: ampicillin (100 μ g/mL), spectinomycin (50 μ g/mL), kanamycin (50 μ g/mL), and chloramphenicol (34 μ g/mL).

[0087] All plasmids used in this study and oligonucleotides used in their construction are listed in Tables 6 and 7. Plasmid based gene overexpression was achieved by cloning

the desired gene(s) into either pETDuet-1 or pCDFDuet-1 (Novagen, Darmstadt, Germany) digested with appropriate restriction enzymes using In-Fusion PCR cloning technology (Clontech Laboratories, Inc., Mountain View, CA). Cloning inserts were created via PCR of ORFs of interest from their respective genomic or codon-optimized DNA with Phusion polymerase (Thermo Scientific, Waltham, MA). *E. coli* genes were obtained from genomic DNA, while heterologous genes were synthesized by GenScript (Piscataway, NJ) or GeneArt (Life Technologies, Carlsbad, CA) with codon optimization except for *bktB*, *phaB1*, and *pct*, which were amplified from genomic DNA or cDNA of their source organisms. The resulting In-Fusion products were used to transform *E. coli* Stellar cells (Clontech Laboratories, Inc., Mountain View, CA) and PCR identified clones were confirmed by DNA sequencing.

TABLE 6

Strains and plasmids used in this study.	
Strain/plasmid	Genotype
<i>E. coli</i> Strains	
MG1655	F- λ -ilvG-rfb-50 rph-1
JC01	MG1655 Δ ldhA::FRT Δ poxB::FRT Δ pta::FRT Δ adhE::FRT Δ frdA::FRT
JC01(DE3)	JC01 with DE3, a λ prophage carrying the T7 RNA polymerase gene and <i>lacI^q</i>
JST06	JC01 Δ ycaA::FRT Δ ybgC::FRT Δ ydhJ::FRT Δ tesA::FRT Δ fadM::FRT Δ tesB::FRT
JST06(DE3)	JST06 with DE3, a λ prophage carrying the T7 RNA polymerase gene and <i>lacI^q</i>

TABLE 6-continued

Strains and plasmids used in this study.	
Strain/plasmid	Genotype
MG1655(DE3)	MG1655 with DE3, a λ prophage carrying the T7 RNA polymerase gene and <i>lacI^q</i>
MG1655(DE3) Δ glcD	MG1655(DE3) Δ glcD::FRT
BL21(DE3)	F- ompT gal dem lon hsdS _B (r _B ⁻ m _B ⁻) λ (DE3 [lacI lacUV5-T7 gene 1 ind1 sam7 nin5]) [malB ⁺] _{K-12} (λ ^S)
<i>S. cerevisiae</i> strains	
INVSc1	MATa his3D1 leu2 trp1-289 ura3-52 MAT his3D1 leu2 trp1-289 ura3-52
Plasmids	
pETDuet	ColE1 (pBR322) ori, lacI, T7lac,
pETDuet-P1-fadB2x-fadB1x	ColE1 ori; Amp ^R ; P _{T7lac-1} : fadB2x-fadB1x
pETDuet-P1-fadB2x-fadB1x-P2-ydil	ColE1 ori; Amp ^R ; P _{T7lac-1} : fadB2x-fadB1x P _{T7lac-2} : ydil
pETDuet-P1-bktB-phaB1	ColE1 ori; Amp ^R ; P _{T7lac-1} : bktB-phaB1
pETDuet-P1-bktB-phaB1-P2-phaJ	ColE1 ori; Amp ^R ; P _{T7lac-1} : bktB-phaB1 P _{T7lac-2} : phaJ
pCDFDuet-1	CloDF13 ori, lacI, T7lac, Strep ^R
pCDFDuet-P1-pct-fadAx	CloDF13 ori; Strep ^R ; P _{T7lac-1} : pct-fadAx
pCDFDuet-P1-pct-fadAx-P2-fabI	CloDF13 ori; Strep ^R ; P _{T7lac-1} : pct-fadAx P _{T7lac-2} : fabI
pCDFDuet-P1-pct-P2-tdTer	CloDF13 ori; Strep ^R ; P _{T7lac-1} : pct P _{T7lac-2} : tdTer
pCDFDuet-1-P1-ntH6-HACL1	CloDF13 ori; Strep ^R ; P _{T7lac-1} : ntHis6-HACL1
pYE260-HACL1	ColE1 ori; Amp ^R ; P _{GAL1} : ntHis6-HACL1

TABLE 7

Oligonucleotides used in this study for plasmid constructions		
Name	SEQ ID	Sequence
pct-f1	1	5' -AGGAGATATACCATGAG AAAAGTAGAAATCATTAC-3'
pct-r1	2	5' -CGCCGAGCTCGAATTCT TATTTTTCAGTCCCATGGG AC-3'
fabI-f1	3	5' -AAGGAGATATACATATG GGTTTCTTTCCGGTAAG-3'
fabI-r1	4	5' -TTGAGATCTGCCATATG TTATTTTCAGTTCGAGTTCGT TC-3'
fadAx-f1	5	5' -GAAAAAATAAGAATTTA AGGAGGAATAAACCATGACC CTGGCAAATGATCC-3'
fadAx-r1	6	5' -CGCCGAGCTCGAATTCT TAATACAGACATCAACTGC C-3'
fadB2x-f1	7	5' -AGGAGATATACCATGCA TATCGCCAACAAACAC-3'
fadB2x-r1	8	5' -CGCCGAGCTCGAATTCT TATTTTGCTGCCATGCGCAG -3'
fadB1x-f1	9	5' -AGCAAAAATAAGAATTTA AGGAGGAATAAACCATGGCC TTTGAAACCATTCTG-3'

TABLE 7-continued

Oligonucleotides used in this study for plasmid constructions		
Name	SEQ ID	Sequence
fadB1x-r1	10	5' -CGCCGAGCTCGAATTCT TAGCGATCTTTAAACTGTGC -3'
ydil-f1	11	5' -AAGGAGATATACATATG ATATGGAAACCGAAAATCAC -3'
ydil-r1	12	5' -TTGAGATCTGCCATATG TCACAAAATGGCGTTCGTC-3'
bktB-f1	13	5' -AGGAGATATACCATGAT GACGCGTGAAGTGGTAGT-3'
bktB-r1	14	5' -CGCCGAGCTCGAATTCT CAGATACGCTCGAAGATGG-3'
phaB1-f1	15	5' -GCGTATCTGAGAATTAG GAGGCTCTCT ATGACTCAG CGCATTGCGTA
phaB1-r1	16	5' -CGCCGAGCTCGAATTCT CAGCCCATGTGCAGGCC-3'
phaJ-f1	17	5' -AAGGAGATATACATATG TCGGCACAAAGCCTG-3'
phaJ-r1	18	5' -TTGAGATCTGCCATATG TTACGGCAGTTTCACCACC-3'

TABLE 7-continued

Oligonucleotides used in this study for plasmid constructions		
Name	SEQ ID	Sequence
HACL1-f1	19	5' -GCCAGGATCCGAATTct ATGCCGGACACCAACTTC-3'
HACL1-r1	20	5' -CGCCGAGCTCGAATTc TTACATATTGCTACGGGTCA GC-3'

[0088] Fermentation medium and conditions: The minimal medium designed by Neidhardt et al. with 125 mM MOPS and Na₂HPO₄ in place of K₂HPO₄ (1.48 mM for fermentations in flasks; 2.8 mM for fermentations in bioreactors), supplemented with 20 g/L glycerol, 10 g/L tryptone, 5 g/L yeast extract, 100 μM FeSO₄, 5 mM calcium pantothenate, 5 mM (NH₄)₂SO₄, and 30 mM NH₄Cl was used for all fermentations unless otherwise stated. Neutralized 20 mM glycolic acid or propionic acid was supplemented as needed. Antibiotics (50 μg/mL carbenicillin and 50 μg/mL spectinomycin) were included when appropriate. All chemicals were obtained from Fisher Scientific Co. (Pittsburg, PA) and Sigma-Aldrich Co. (St. Louis, MO).

[0089] Unless otherwise stated, fermentations were performed in 25 mL Pyrex Erlenmeyer flasks (narrow mouth/heavy duty rim, Corning Inc., Corning, NY) filled with 20 mL fermentation medium and sealed with foam plugs filling the necks. A single colony of the desired strain was cultivated overnight (14-16 h) in LB medium with appropriate antibiotics and used as the inoculum (1%). After inoculation, flasks were incubated in a NBS I24 Benchtop Incubator Shaker (New Brunswick Scientific Co., Inc., Edison, NJ) at 200 rpm and 37° C., except fermentations supplemented with phenylacetic acid or isobutyric acid in which the temperature was 30° C. When optical density (550 nm, OD550) reached ~0.3-0.5, 5 μM isopropyl β-d-1-thiogalactopyranoside (IPTG) was added for plasmid based gene expression in all cases except the following: 1 μM IPTG was used for adipic acid production from glycerol without succinic acid supplementation and 10 μM IPTG was used during production of ω-phenylalkanoic acids. For induction of controlled chromosomal expression constructs, 0.1 mM cumate and 15 ng/mL anhydrotetracycline were also added when appropriate. Flasks were then incubated under the same conditions for 48 h post-induction unless otherwise stated.

[0090] Additional fermentations were conducted in a Six-Fors multi-fermentation system (Infors HT, Bottmingen, Switzerland) with an air flow rate of 2 N L/hr, independent control of temperature (37° C.), pH (controlled at 7.0 with NaOH and H₂SO₄), and stirrer speed (720 rpm). Tiglic acid fermentations used the previously described fermentation media with 30 g/L glycerol, the inclusion of 5 μM sodium selenite, and 5 μM IPTG. Propionic acid (20 mM) was added at 0, 24, and 48 h. Pre-cultures were grown in 25 mL flasks as described above, incubated for 4 h post-induction, and used for inoculation as described above.

[0091] Fermentations with glycolyl-CoA as a primer were conducted in 250 mL Erlenmeyer Flasks filled with 50 mL LB media supplemented with 10 g/L glucose and appropriate antibiotics. The cultivation of inoculum was same as above but 2% inoculation was used. After inoculation, cells

were cultivated at 30° C. and 250 rpm in a NBS 124 Benchtop Incubator Shaker until an optical density of ~0.8 was reached, at which point IPTG (0.1 mM) and neutralized glycolic acid (40 mM) were added. Flasks were then incubated under the same conditions for 96 h post induction.

[0092] GC sample preparation: Sample preparation was conducted as follows: 2 mL culture supernatant samples were transferred to 5 mL glass vials (Fisher Scientific Co., Fair Lawn, NJ, USA) and 80 μL of 50% H₂SO₄ and 340 μL of 30% NaCl solution were added for pH and ionic strength adjustment, respectively. Tridecanoic acid (final concentration 50 mg/L) was added as internal standard and 2 mL of hexane-MTBE (1:1) added for extraction. The bottles were sealed with Teflonlined septa (Fisher Scientific Co., Fair Lawn, NJ, USA), secured with caps, and rotated at 60 rpm for 120 min. The samples were then centrifuged for 2 min at 2,375×g to separate the aqueous and organic layers. 1 mL of the dry organic layer was transferred into a 2 mL borosilicate glass vial, dried under N₂, and re-suspended in 1000 μL of pyridine. After vortexing, 100 μL of BSTFA (N,O-bis(trimethylsilyl)trifluoroacetamide) was added, the samples were heated at 70° C. for 30 min, dried under N₂ and re-suspended in 1 mL hexane for analysis.

[0093] GC-MS metabolite identification: Except identifications of 2,3-dihydroxybutyric acid, metabolite identification was conducted via GC-MS in an Agilent 7890A GC system (Agilent Technologies, Santa Clara, CA), equipped with a 5975C inert XL mass selective detector (Agilent) and Rxi-5Sil column (0.25 mm internal diameter, 0.10 μm film thickness, 30 m length; Restek, Bellefonte, PA). The sample injection amount was 2 μL with 40:1 split ratio. The injector and detector were maintained at 280° C. The column temperature was held initially at 35° C. for 1 min and increased to 200° C. at the rate of 6° C./min, then to 270° C. at the rate of 30° C./min. That final temperature was maintained for 1 min before cooling back to initial temperature. The carrier gas was helium (2.6 mL/min, Matheson Tri-Gas, Longmont, CO).

[0094] Identification of 2,3-dihydroxybutyric acid was conducted by the Baylor College of Medicine Analyte Center (bcm.edu/research/centers/analyte, Houston, TX). An Agilent 6890 GC system (Agilent Technologies, Santa Clara, CA), equipped with a 5973 mass selective detector (Agilent Technologies) and HP-5 ms column (Agilent Technologies) was used. Sample extraction was conducted using Agilent Chem Elut liquid extraction columns (Agilent Technologies) according to manufacturer protocols.

[0095] HPLC metabolite quantification: The concentration of products were determined via ion-exclusion HPLC using a Shimadzu Prominence SIL 20 system (Shimadzu Scientific Instruments, Inc., Columbia, MD) equipped with an HPX-87H organic acid column (Bio-Rad, Hercules, CA) with operating conditions to optimize peak separation (0.3 ml/min flow rate, 30 mM H₂SO₄ mobile phase, column temperature 42° C.).

[0096] In vitro enzyme assay: Purified HACL1 was tested for its native catabolic activity by assessing its ability to cleave 2-hydroxyhexadecanoyl-CoA to pentadecanal and formyl-CoA. Enzyme assays were performed in 50 mM tris-HCl pH 7.5, 0.8 mM MgCl₂, 0.02 mM TPP, 6.6 μM BSA, and 0.3 mM 2-hydroxyhexadecanoyl-CoA. The assay mixtures were incubated for one hour at 37° C., after which the presence of pentadecanal was assessed by extraction with hexane and analysis by GC-FID.

[0097] 2-hydroxyhexadecanoyl-CoA was prepared by the n-hydroxysuccinimide method. In summary, the n-hydroxysuccinimide ester of 2-hydroxyhexadecanoic acid is prepared by reacting n-hydroxysuccinimide with the acid in the presence of dicyclohexylcarbodiimide. The product was filtered and purified by recrystallization from methanol to give pure n-hydroxysuccinimide ester of 2-hydroxyhexadecanoic acid. The ester was reacted with CoA-SH in presence of thioglycolic acid to give 2-hydroxyhexadecanoyl-CoA. The 2-hydroxyhexadecanoyl-CoA was purified precipitation using perchloric acid, filtration, and washing the filtrate with perchloric acid, diethyl ether, and acetone.

[0098] For specific activity assays (reported in μmol substrate/mg protein/min) these supernatant fractions were utilized and protein concentration was established using the Bradford Reagent (Thermo Sci.) using BSA as the protein standard.

[0099] Enzyme purification: A plasmid containing the codon optimized gene encoding human HIS-tagged HAC1 was constructed as described. The resulting construct was transformed into *S. cerevisiae* InvSC1 (Life Tech.). The resulting strain was cultured in 50 mL of SC-URA media containing 2% glucose at 30° C. for 24 hours. The cells were pelleted and the required amount of cells were used to inoculate a 250 mL culture volume of SC-URA media containing 0.2% galactose, 1 mM MgCl_2 , and 0.1 mM thiamine to 0.4 OD600. After 20 hours incubation with shaking at 30° C., the cells were pelleted and saved.

[0100] When needed, the cell pellets were resuspended to an OD600 of approximately 100 in a buffer containing 50 mM potassium phosphate pH 7.4, 0.1 mM thiamine pyrophosphate, 1 mM MgCl_2 , 0.5 mM AEBSEF, 10 mM imidazole, and 250 units of Benzoylase nuclease. To the cell suspension, approximately equal volumes of 425-600 μm glass beads were added. Cells were broken in four cycles of 30 seconds of vortexing at 3000 rpm followed by 30 seconds on ice. The glass beads and cell debris were pelleted by centrifugation and supernatant containing the cell extract was collected. The HIS-tagged HAC1 was purified from the cell extract using Talon Metal Affinity Resin as described above, with the only modification being the resin bed volume and all subsequent washes were halved. The eluate was collected in two 500 μL fractions.

[0101] Expression and purification of the desired protein can be confirmed by running cell pellet sample and eluate on SDS-PAGE.

[0102] We demonstrated several cases of the iterative system can synthesize alpha-functionalized small molecules through the use of alpha-functionalized forms of acetyl-CoA as the extender unit. One case used of propionyl-CoA as the extender unit. To implement this, *P. putida* FadAx (thiolase), FadB2x (HACD), FadB1x (ECH), and *E. coli* FabI (ECR) were used with Pct for activation of exogenous propionic acid. Expression in JC01(DE3) resulted in the production of 2-methylbutyric acid (75 mg/L) and tiglic acid (573 mg/L) (FIG. 5), representing products of acid-forming endogenous termination enzymes at the acyl-CoA and enoyl-CoA pathway nodes.

[0103] Interestingly, 2-methylpentanoic acid (49 mg/L) and (E)-2-methyl-2-pentenoic acid (84 mg/L) were also synthesized, as the result of propionyl-CoA serving as both the primer and the extender unit. Products resulting from non-functionalized extender units (acetyl-CoA) with acetyl-CoA or propionyl-CoA priming were also observed, demonstrating the nonspecific activity of the thiolase (and subsequent β -reduction enzymes). This represents a potential area for further improvement through the selection and engineering of a thiolase with maximal specificity for the desired condensation. Additional alpha-functionalization was demonstrated with glycolyl-CoA (i.e. α -hydroxylated acetyl-CoA) as the extender unit, which with acetyl-CoA priming supported the synthesis of 2,3-dihydroxybutyric acid (FIG. 11).

[0104] The ability of the alpha-functionalization system to support high product titers was investigated by improving tiglic acid production. Omission of ECR and manipulation of the termination pathway through deletion of native thioesterases and controlled overexpression of Ydi1, a thioesterase previously shown to act effectively on α,β -unsaturated enoyl-CoAs, resulted in further improvement, from 573 mg/L to 1.39 g/L (FIG. 7). When a controlled bioreactor with a higher initial glycerol concentration was used, tiglic acid production increased to 3.79 g/L (11.6% mol/mol glycerol) (FIG. 8).

[0105] The host strains and plasmids used for production of above products are summarized in Table 8.

TABLE 8

Host strains and plasmids enabling alpha-functionalized small molecule synthesis with listed primer/extender unit combinations					
Host strain	Plasmid 1	Plasmid 2	Primer	Extender unit	Product
JC01(DE3)	pETDuet-P1-fadB2x-fadB1x	pCDFDuet-P1-pct-fadAx-P2-fabI	Acetyl-CoA	Propionyl-CoA	2-methylbutyric acid Tiglic acid
			Propionyl-CoA	Propionyl-CoA	2-methylpentanoic acid (E)-2-methyl-2-pentenoic acid
JC01(DE3)	pETDuet-P1-fadB2x-fadB1x	pCDFDuet-P1-pct-fadAx	Acetyl-CoA	Propionyl-CoA	Tiglic acid
JST06(DE3)	pETDuet-P1-fadB2x-fadB1x	pCDFDuet-P1-pct-fadAx	Acetyl-CoA	Propionyl-CoA	N.A.
JST06(DE3)	pETDuet-P1-fadB2x-fadB1x	pCDFDuet-P1-pct-fadAx	Acetyl-CoA	Propionyl-CoA	Tiglic acid
		P2-ydi1			

TABLE 8-continued

Host strains and plasmids enabling alpha-functionalized small molecule synthesis with listed primer/extender unit combinations					
Host strain	Plasmid 1	Plasmid 2	Primer	Extender unit	Product
Acetyl-CoA	Glycolyl-CoA	2,3-dihydroxybutyric acid	Acetyl-CoA	Glycolyl-CoA	2,3-dihydroxybutyric acid

[0106] We also successfully expressed *Homo sapiens* 2-hydroxyacyl-CoA lyase HAC1 in *Saccharomyces cerevisiae* and *Escherichia coli* (FIGS. 14 and 16), and confirmed its activity of degradation of 2-hydroxyhexadecanoyl-CoA to pentadecanal (FIGS. 17-18). This provides the potential of combination of 2-hydroxyacyl-CoA lyase with proposed iterative platform using alpha-hydroxylated glycolyl-CoA as the extender unit for the synthesis of primary alcohols.

[0107] We believe that, pathway and process optimization, in line with industrial biotechnology approaches, can further improve performance for a specific target product, as the underlying carbon and energy efficiency enables the feasibility of further advancing product titer, rate, and yield. Important areas include generating and balancing pools of priming and extender units and optimization of required pathway enzymes for a given target product. The former can exploit previously developed pathways for primers and extender units, whereas the latter includes identifying and engineering enzymes that may be flux limiting due to suboptimal enzyme specificity or activity. These approaches will be continually aided by developments in protein and metabolic engineering and synthetic and systems biology.

[0108] The above experiments are repeated in *Bacillus subtilis*. The same genes can be used, especially since *Bacillus* has no significant codon bias. A protease-deficient strain like WB800N is preferably used for greater stability of heterologous protein. The *E. coli*-*B. subtilis* shuttle vector pMTLBS72 exhibiting full structural stability can be used to move the genes easily to a more suitable vector for *Bacillus*. Alternatively, two vectors pHT01 and pHT43 allow high-level expression of recombinant proteins within the cytoplasm. As yet another alternative, plasmids using the theta-mode of replication such as those derived from the natural plasmids pAM β 1 and pBS72 can be used. Several other suitable expression systems are available. Since the FAS genes are ubiquitous, the invention is predicted to function in *Bacillus*.

[0109] The above experiments are repeated in yeast. The same genes can be used, but it may be preferred to accommodate codon bias. Several yeast *E. coli* shuttle vectors are available for ease of the experiments. Since the FAS genes are ubiquitous, the invention is predicted to function in yeast, especially since yeasts are already available with exogenous functional TE genes and the reverse beta oxidation pathway has also been made to run in yeast.

[0110] Each of the following is incorporated by reference herein in its entirety for all purposes:

[0111] U.S. 20130316413 Reverse beta oxidation pathway.

[0112] 62/140,628 Bioconversion of short-chain hydrocarbons to fuels and chemicals, Mar. 31, 2015.

[0113] WO2015112988 Type II fatty acid synthesis enzymes in reverse beta-oxidation, Jan. 26, 2015 and 61/932,057, Jan. 27, 2014.

[0114] 62/069,850 Synthetic pathway for biosynthesis from 1-carbon compounds, Oct. 29, 2014.

[0115] 61/531/911, Sep. 7, 2011; 61/440,192, Feb. 7, 2011, U.S. 20140273110, WO2013036812 Functionalized carboxylic acids and alcohols by reverse fatty acid oxidation.

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[0122] Clomburg, J. M., et al., Synthetic Biology Approach to Engineer a Functional Reversal of the β -Oxidation Cycle. *ACS Synthetic Biology* 1, 541-554 (2012).

[0123] Vick, J. E. et al. *Escherichia coli* enoyl-acyl carrier protein reductase (FabI) supports efficient operation of a functional reversal of the β -oxidation cycle. *Appl. Environ. Microbiol.* 81, 1406-1416 (2015).

[0124] Cheong, S., Clomburg, J. M. and Gonzalez, R.* (2016). Energy- and carbon-efficient synthesis of functionalized small molecules in bacteria using non-decarboxylative Claisen condensation reactions. *Nat. Biotechnol.* 34 (5): doi:10.1038/nbt.3505.

[0125] The following claims are provided to add additional clarity to this disclosure. Future applications claiming priority to this application may or may not include the following claims, and may include claims broader, narrower, or entirely different from the following claims. Further, any detail from any claim may be combined with any other detail from another claim, even if not yet so combined.

SEQUENCE LISTING

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source 1..35

mol_type = other DNA

organism = synthetic construct

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FEATURE Location/Qualifiers

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note = Synthetic

source 1..39

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FEATURE Location/Qualifiers

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mol_type = other DNA

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FEATURE Location/Qualifiers

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                  organism = synthetic construct

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source            1..32
                  mol_type = other DNA
                  organism = synthetic construct

SEQUENCE: 17
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source            1..36
                  mol_type = other DNA
                  organism = synthetic construct

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                  mol_type = other DNA
                  organism = synthetic construct

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SEQ ID NO: 20      moltype = DNA length = 38
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                  note = Synthetic
source            1..38
                  mol_type = other DNA
                  organism = synthetic construct

SEQUENCE: 20
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1. A genetically engineered bacteria, said bacteria comprising:

- a) an overexpressed acyl-CoA transferase enzyme that converts glycolate to glycolyl-CoA or propionate to propionyl-CoA and wherein said overexpressed acyl-CoA transferase enzyme is encoded by a *Megasphaera elsdenii* pct gene;
- b) an overexpressed thiolase enzyme that catalyzes a condensation of an acetyl-CoA primer or a propionyl-CoA primer with a propionyl-CoA extender or an acetyl-CoA primer with glycolyl-CoA extender to generate 2-methyl-3-ketobutyryl-CoA or 2-methyl-3-ketopentanoyl-CoA or 2-hydroxy-3-ketobutyryl-CoA, wherein said overexpressed thiolase enzyme is encoded by a gene(s) selected from a group consisting of *Pseudomonas putida* fadAx, *Ralstonia eutropha* bktB, *Escherichia coli* atoB, *Escherichia coli* yqeF, *Escherichia coli* fadA, *Escherichia coli* fadI, *Pseudomonas* sp. B13 catF, *Escherichia coli* paaJ, *Rhodococcus opacus* pcaF, *Pseudomonas putida* pcaF, *Streptomyces* sp.

- c) an overexpressed 3-hydroxyacyl-CoA dehydrogenase enzyme that catalyzes a reduction of 2-methyl-3-ketobutyryl-CoA to 2-methyl-3-hydroxybutyryl-CoA or 2-methyl-3-ketopentanoyl-CoA to 2-methyl-3-hydroxypentanoyl-CoA or 2-hydroxy-3-ketobutyryl-CoA to 2,3-dihydroxybutyryl-CoA, wherein said overexpressed 3-hydroxyacyl-CoA dehydrogenase enzyme is encoded by a gene(s) selected from a group consisting of *Pseudomonas putida* fadB2x, *Ralstonia eutropha* phaB, *Escherichia coli* fadB, *Escherichia coli* fadJ, *Escherichia coli* paaH, *Pseudomonas putida* fadB, *Acinetobacter* sp. ADP1 dcaH, and *Clostridium acetobutylicum* hbd;
- d) an enoyl-CoA hydratase or a 3-hydroxyacyl-CoA dehydratase enzyme that catalyzes a dehydration of 2-methyl-3-hydroxybutyryl-CoA to 2-methyl-crotonoyl-CoA or 2-methyl-3-hydroxypentanoyl-CoA to

- 2-methyl-pentenoyl-CoA or 2,3-dihydroxybutyryl-CoA to 2-hydroxy-crotonoyl-CoA, wherein said over-expressed enoyl-CoA hydratase or 3-hydroxyacyl-CoA dehydratase enzyme is encoded by a gene(s) selected from a group consisting of *Pseudomonas putida* fadB1x, *Escherichia coli* fabA, *Escherichia coli* fabZ, *Escherichia coli* fadB, *Escherichia coli* fadJ, *Escherichia coli* paaF, *Pseudomonas putida* fadB, *Acinetobacter* sp. ADP1 dcaE, *Clostridium acetobutylicum* crt, and *Aeromonas caviae* phaJ;
- e) a trans-enoyl-CoA reductase or an enoyl-[acyl-carrier-protein]reductase enzyme that catalyzes a reduction of 2-methyl-crotonoyl-CoA to 2-methyl-butyryl-CoA or 2-methyl-pentenoyl-CoA to 2-methyl-pentanoyl-CoA or 2-hydroxy-crotonoyl-CoA to 2-hydroxy-butyryl-CoA, wherein said trans-enoyl-CoA reductase or enoyl-[acyl-carrier-protein] reductase enzyme is encoded by a gene(s) selected from a group consisting of *Escherichia coli* fabI, *Euglena gracilis* TER, *Treponema denticola* TER, *Clostridium acetobutylicum* TER, *Enterococcus faecalis* fabK, *Bacillus subtilis* fabL, and *Vibrio cholerae* fabV;
- f) a termination enzyme(s) able to use a substrate selected from any CoA thioester intermediate produced by enzymes b-e to make a product selected from 2-methyl-3-ketobutyrate, 2-methyl-3-hydroxybutyrate, 2-methyl-3-ketopentanoate, 2-methyl-3-hydroxypentanoate, 2-hydroxy-3-ketobutyrate, 2,3-dihydroxybutyrate, 2-methyl-crotonoate, 2-methyl-butyrate, 2-methyl-pentenoate, 2-methyl-pentanoate, 2-hydroxy-crotonoate, or 2-hydroxy-butyrate, wherein said termination enzyme(s) is selected from a group consisting of:
- i) a thioesterase encoded by a gene(s) selected from a group consisting of *Escherichia coli* tesA, *Escherichia coli* tesB, *Escherichia coli* yciA, *Escherichia coli* fadM, *Escherichia coli* ydiI, *Escherichia coli* ybgC, *Escherichia coli* paaI, *Mus musculus* acot8, *Alcanivorax borkumensis* tesB2, *Fibrobacter succinogenes* Fs2108, *Prevotella ruminicola* Pr655, *Prevotella ruminicola* Pr1687, and *Lycopersicon hirsutum* f. *glabratum* mks2;
- ii) an acyl-CoA transferase encoded by a gene(s) selected from a group consisting of *Escherichia coli* atoD, *Escherichia coli* scpC, *Escherichia coli* ydiF, *Escherichia coli* atoA, *Escherichia coli* atoD, *Clostridium acetobutylicum* ctfA, *Clostridium acetobutylicum* ctfB, *Clostridium kluyveri* cat2, *Clostridium kluyveri* cat1, *Pseudomonas putida* pcal, *Pseudomonas putida* pcaJ, *Megasphaera elsdenii* pct, *Acidaminococcus fermentans* gctA, *Acidaminococcus fermentans* gctB, and *Acetobacter aceti* aarC; or
- iii) a phosphotransacylase encoded by a gene(s) selected from a group consisting of *Clostridium acetobutylicum* ptb, *Enterococcus faecalis* ptb, and *Salmonella enterica* pduL and a carboxylate kinase encoded by a gene(s) selected from a group consisting of *Clostridium acetobutylicum* buk, *Enterococcus faecalis* buk, and *Salmonella enterica* pduW.
2. A genetically engineered bacteria, said bacteria comprising:
- a *Megasphaera elsdenii* pct gene encoding acyl-CoA transferase enzyme that converts propionate to propionyl-CoA;
 - a *Pseudomonas putida* fadAx gene encoding a thiolase that catalyzes the condensation of an acetyl-CoA primer or a propionyl-CoA primer with a propionyl-CoA extender to generate 2-methyl-3-ketobutyryl-CoA or 2-methyl-3-ketopentanoyl-CoA;
 - a *Pseudomonas putida* fadB2x encoding a 3-hydroxyacyl-CoA dehydrogenase enzyme that catalyzes a reduction of 2-methyl-3-ketobutyryl-CoA to 2-methyl-3-hydroxybutyryl-CoA or 2-methyl-3-ketopentanoyl-CoA to 2-methyl-3-hydroxypentanoyl-CoA;
 - a *Pseudomonas putida* fadB1x encoding an enoyl-CoA hydratase or a 3-hydroxyacyl-CoA dehydratase enzyme that catalyzes a dehydration of 2-methyl-3-hydroxybutyryl-CoA to 2-methyl-crotonoyl-CoA or 2-methyl-3-hydroxypentanoyl-CoA to 2-methyl-pentenoyl-CoA;
 - an *Escherichia coli* fabI gene encoding a trans-enoyl-CoA reductase or an enoyl-[acyl-carrier-protein] reductase enzyme that catalyzes a reduction of 2-methyl-crotonoyl-CoA to 2-methyl-butyryl-CoA or 2-methyl-pentenoyl-CoA to 2-methyl-pentanoyl-CoA;
 - an *Escherichia coli* ydiI gene encoding a thioesterase enzyme that catalyzes a conversion of 2-methyl-crotonoyl-CoA to 2-methyl-crotonoate or 2-methyl-butyryl-CoA to 2-methyl-butyrate or 2-methyl-pentenoyl-CoA to 2-methyl-pentenoate or 2-methyl-pentanoyl-CoA to 2-methyl-pentanoate.
3. A genetically engineered bacteria, said bacteria comprising:
- a *Megasphaera elsdenii* pct gene encoding acyl-CoA transferase enzyme that converts propionate to propionyl-CoA;
 - a *Ralstonia eutropha* bktB gene encoding a thiolase that catalyzes the condensation of an acetyl-CoA primer with glycolyl-CoA extender to generate 2-hydroxy-3-ketobutyryl-CoA;
 - a *Ralstonia eutropha* phaB gene encoding a 3-hydroxyacyl-CoA dehydrogenase enzyme that catalyzes a reduction of 2-hydroxy-3-ketobutyryl-CoA to 2,3-dihydroxybutyryl-CoA;
 - an endogenous *Escherichia coli* gene encoding a thioesterase enzyme that catalyzes the conversion of 2,3-dihydroxybutyryl-CoA to 2,3-dihydroxybutyrate.
4. A method of producing an alpha functionalized product, comprising growing the bacteria of claim 1 in a nutrient broth containing an alpha functionalized acid selected from propionic acid or glycolic acid under conditions such that said gene(s) are expressed, said microorganism producing an alpha functionalized product from said alpha-functionalized acid by reverse beta oxidation, wherein said alpha functionalized product is tiglic acid, 2-methylbutyric acid, 2-methylpentenoic acid, 2-methyl-2-pentenoic acid or 2-methylvaleric acid.
5. A method of making an alpha functionalized product, comprising growing the bacteria of claim 1 in a nutrient broth under conditions such that said gene(s) are expressed, said microorganism producing an alpha functionalized product from an alpha-functionalized acid by reverse beta oxidation, and isolating said alpha functionalized product.
6. The method of claim 5, wherein said nutrient broth is supplemented with said alpha-functionalized acid.

7. The method of claim 6, wherein said alpha-functionalized acid is propionic acid or glycolic acid.

8. The method of claim 7, wherein said alpha functionalized product is tiglic acid, 2-methylbutyric acid, 2-methylpentenoic acid, 2-methyl-2-pentenoic acid or 2-methylvaleric acid.

9. A method of producing an alpha functionalized product, comprising growing the bacteria of claim 2 in a nutrient broth containing an alpha functionalized acid selected from propionic acid or glycolic acid under conditions such that said gene(s) are expressed, said microorganism producing an alpha functionalized product from said alpha-functionalized acid by reverse beta oxidation, wherein said alpha functionalized product is selected from 2-methyl-butyrate, 2-methyl-pentanoate, 2-methyl-crotonoate, 2-methyl-pentenoate, and 2,3-dihydroxybutyrate.

10. A method of making an alpha functionalized product, comprising growing the bacteria of claim 2 in a nutrient broth under conditions such that said gene(s) are expressed, said microorganism producing an alpha functionalized product from an alpha-functionalized acid by reverse beta oxidation, and isolating said alpha functionalized product or a beta- or omega-functionalized derivative thereof.

11. The method of claim 10, wherein said nutrient broth is supplemented with said alpha-functionalized acid.

12. The method of claim 11, wherein said alpha-functionalized acid is propionic acid or glycolic acid.

13. The method of claim 12, wherein said alpha functionalized product is 2-methyl-butyrate, 2-methyl-pentanoate, 2-methyl-crotonoate, 2-methyl-pentenoate, and 2,3 -dihydroxybutyrate.

14. A method of producing an alpha functionalized product, comprising growing the bacteria of claim 3 in a nutrient broth containing an alpha functionalized acid selected from propionic acid or glycolic acid under conditions such that said gene(s) are expressed, said microorganism producing an alpha functionalized product from said alpha-functionalized acid by reverse beta oxidation, wherein said alpha functionalized product is selected from 2-methyl-butyrate, 2-methyl-pentanoate, 2-methyl-crotonoate, 2-methyl-pentenoate, and 2,3-dihydroxybutyrate.

15. A method of making an alpha functionalized product, comprising growing the bacteria of claim 3 in a nutrient broth under conditions such that said gene(s) are expressed, said microorganism producing an alpha functionalized product from an alpha-functionalized acid by reverse beta oxidation, and isolating said alpha functionalized product or a beta- or omega-functionalized derivative thereof.

16. The method of claim 15, wherein said nutrient broth is supplemented with said alpha-functionalized acid.

17. The method of claim 16, wherein said alpha-functionalized acid is propionic acid or glycolic acid.

18. The method of claim 17, wherein said alpha functionalized product 2-methyl-butyrate, 2-methyl-pentanoate, 2-methyl-crotonoate, 2-methyl-pentenoate, and 2,3-dihydroxybutyrate.

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